Package 'PGSEA'

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Type Package

Title Parametric Gene Set Enrichment Analysis

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Description Parametric Analysis of Gene Set Enrichment

Depends R (>= 2.10), GO.db, KEGG.db, AnnotationDbi, annaffy, methods,
Biobase (>= 2.5.5)

Suggests GSEABase, GEOquery, org.Hs.eg.db, hgu95av2.db, limma

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ZipData no

biocViews Microarray

LazyData no

NeedsCompilation no

R topics documented:

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  aggregateExprs   *Aggregate expression data*

Description

This function removes duplicates row names from an expression set, summarizing them with a function of the users choice. The "absMax" function located in package "reb" we have found to be useful.

Usage

```
aggregateExprs(x, package = "hgu133plus2", using = "ENTREZID", FUN, ...)
```

Arguments

x	expression data - matrix, eSet, or ExpressionSet
package	annotation package of expression data
using	format type that gene IDs are converted to
FUN	function by which to summarize duplicated values
...	extra parameters passed on to FUN

Value

A matrix of expression data with the rows aggregated to a unique format chosen by the user. The new identifiers of the returned matrix are those specified with the "using" argument. To see possible values, use the ls() command illustrated below in the examples.

Author(s)

Kyle Furge <kyle.furge@vai.org> and Karl Dykema <karl.dykema@vai.org>

Examples

```
if (require(hgu95av2.db) & require(annaffy)) {
library(annaffy)
data(aafExpr)
class(exprs(aafExpr))
exprs(aafExpr)[1:4, 1:4]

#list possible values for the "using" argument
ls(pos=which(search()=="package:hgu95av2.db"))
```

```
convert <- aggregateExprs(exprs(aafExpr),"hgu95av2.db",FUN=mean,na.rm=TRUE)
convert[1:4,1:4]
}
```

---

convertSmc   *Convert Entrez ID based "smc" object*

Description

This function will convert the Entrez IDs of an smc object to the corresponding Entrez IDs from a different species. Data from the homologene project is downloaded and used within this function.

Usage

```
convertSmc(mcs, fromSpecies = "h", toSpecies = "r",hgX="./homologene.data")
```

Arguments

mcs   a list of "smc" objects

fromSpecies   character - a single letter describing the species to convert from ie, h=human, r= rat, etc..

toSpecies   character - a single letter describing the species to convert to ie, h=human, r= rat, etc..

hgX   character - file name of homologene data file

Details

This function will not work if you have not downloaded the homologene data file. Please use this command to do so: download.file("ftp://ftp.ncbi.nih.gov/pub/HomoloGene/current/homologene.data",destfile="homologene.

Value

a list of converted "smc" objects

Author(s)

Karl Dykema <karl.dykema@vai.org>

Examples

```
## Not run:
download.file("ftp://ftp.ncbi.nih.gov/pub/HomoloGene/current/homologene.data",destfile="homologene.data",mode=

datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))

converted <- convertSmc(sample[1:2],"h","r")
```

```
str(converted)

## End(Not run)
```

---

editSmc   *Edit "smc" objects*

Description

This function will edit a single or list of "smc" objects.

Usage

```
editSmc(smcList, attName = "creator", newAtt = "changed!!")
```

Arguments

smcList   a list of "smc" objects

attName   character - which slot to change

newAtt   character - what to change the slot to

Value

a list of edited "smc" objects

Author(s)

Karl Dykema <karl.dykema@vai.org

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
str(sample[1:2])

temp <- editSmc(sample[1:2],"creator","Joe Smith")

str(temp)
```

---

go2smc   *Gene Ontology 2 "smc"*

Description

This function creates "smc" objects from the "GO" Bioconductor library.

Usage

```
go2smc(min = 50, max = 200,organism="human")
```

Arguments

min	numeric - minimum length of ids to be included
max	numeric - maximum length of ids to be included
organism	character - organism

Value

a list of "smc" objects

Author(s)

Karl Dykema <karl.dykema@vai.org>

Examples

```
if(require(GO.db)){
mcs <- go2smc()[1:2]
str(mcs)
}
```

GOLUBmcs   *Molecular Concepts prepared at VAI from data created by Golub et al.*

Description

386 molecular concepts generated at VAI. The data these concepts were generate from is available from <http://www.broad.mit.edu/cmap/>.

Usage

```
data(GOLUBmcs)
```

Format

a list of "smc" objects

Details

These concepts were generated using the limma BioConductor package. The code used for genera-
tion of these concepts is available upon request.

Source

<http://www.broad.mit.edu/cmap/>

Examples

```
data(GOLUBmcs)
str(GOLUBmcs[1:4])
```

---

kegg2smc   *KEGG pathway to "smc"*

Description

This function creates "smc" objects from the "KEGG" Bioconductor library.

Usage

```
kegg2smc(min = 1, max = 284,organism="human")
```

Arguments

min   numeric - minimum length of ids to be included

max   numeric - maximum length of ids to be included

organism   character - organism

Value

a list of "smc" objects

Author(s)

Karl Dykema <karl.dykema@vai.org> and Richard Birnie <richard.birnie@pro-curetherapeutics.com>

Examples

```
if(require(KEGG.db)){
mcs <- kegg2smc(min=20,max=284)
length(mcs)
str(mcs[[1]])
}
```

---

nbEset   *Reduced Neuroblastoma data set*

Description

Neuroblastoma Data set - reduced in size to comply with BioC package guidelines

Usage

```
data(nbEset)
```

Details

This dataset was retrieved from GEO http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE3960 and consists of five reference samples, and ten primary neuroblastoma tumors. Four of the five reference samples GSM2827, GSM2842, GSM2883, and GSM2895 came from a separate dataset, http://www.ncbi.nlm.nih.gov/geo/gds/gds_browse.cgi?gds=181

Source

http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE3960

Examples

```
data(nbEset)
nbEset
```

PGSEA   *Parametric Gene Set Enrichment Analysis*

Description

This package contains functions for an exploratory parametric analysis of gene expression data. This type of analysis can assist in determining of lists of genes, such as those deregulated in defined experimental systems, are similarly deregulated in other data sets.

This function subsets the data based on lists of genes, computes a summary statistic for each gene list, and returns the results in a convenient form.

Usage

```
PGSEA(exprs, cl, range = c(25, 500), ref = NULL, center = TRUE, p.value = 0.005, weighted = TRUE, enfor
```

Arguments

exprs	matrix expression data, a numeric matrix, eSet, or ExpressionSet
cl	gene set list - "GeneSetCollection" or list of "SMC" objects
range	a 2 element vector describing the min and max length of concepts to analyze
enforceRange	boolean - if TRUE, the expression matrix must contain data for the proper number of genes as set by the range argument to return a significant result. (this argument is used for data that contains NA's...)
ref	a vector containing the index of reference samples from which to make comparisons. Defaults to NULL (internally referenced samples)
center	boolean - median center gene expression matrix columns prior to analysis. Can be helpful if 'ref' is used
p.value	numeric p.value threshold or NA to return all data or TRUE to return a matrix of p.values
weighted	boolean - weight results by the size of each gene list
...	extra arguments passed along to FUN

Details

Gene expression values are separated into subsets based on the lists of genes contained in the cl argument. This can be a "GeneSetCollection" or a list of "SMC" (Simple Molecular Concept) objects. For example, readGmt can be used to produce a 'smc' object list from a simple tab-delimited text file. The gene expression values from each of these gene lists is extracted and a summary statistic is computed for each subset (or region in the case of chromosomal bands/arms).

The expression data must have the same identifiers as the list of genes being tested. If they are not, the expression data can be converted using the aggregateExprs function, that can use a current annotation environment to convert and condense the gene expression data.

By default the method set out by Kim and Volsky http://www.biomedcentral.com/1471-2105/6/144 is applied to the gene set. If weighted==FALSE than the default t.test function is used.

The function is set up to perform the analysis on individual samples. For convenient method to analyze groups of samples, see the "Limma User's Guide" for more information on how to see up a contrast matrix and perform a linear model fit. The coefficients of the fit can then be used a input into the PGSEA function.

This package has not been extensively tested beyond a set of well defined curated pathways using the Affymetrix platform and significance values represent approximations. Any results should be confirmed by additional gene set testing methodologies.

Value

If p.value is set to a number, a matrix of results that pass at that significance is returned, of size <number of samples> x <number of molecular concepts>.

If p.value is set to NA, all results are returned.

If p.value is set to TRUE, then a list is returned that consists of the PGSEA results as well as their p.values.

Note

<http://www.biomedcentral.com/1471-2105/6/144>

Author(s)

Kim SY, Volsky DJ., kyle.furge@vai.org and karl.dykema@vai.org

References

PGSEA: Parametric Analysis of Gene Set Enrichment

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
data(nbEset)
pg <- PGSEA(nbEset,cl=sample,ref=1:5)

print(pg[,-c(1:5)])
```

---

readGmt   *readGmt*

Description

This function will read a "gmt" file into R, returning results as a list of SMC objects.

Usage

```
readGmt(fname)
```

Arguments

fname   File name of concepts in .gmt format

Details

The .gmt file format is a tab delimited file format used to store gene lists. These gene lists are stored row by row. The first column is the gene set name. The second column is a brief description, and every entry after that is a gene within that gene set.

Value

A list of SMC objects

Author(s)

Karl Dykema <karl.dykema@vai.org>

References

http://www.broad.mit.edu/gsea/doc/data_formats.html#gmt

See Also

writeGmt

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
str(sample)
```

---

readSmc   *Read SMC files*

Description

This function reads in SMCs (simple molecular concepts) from individual text files.

Usage

```
readSmc(files)
```

Arguments

files   a character vector of file names

Value

A list of SMC objects

Author(s)

Kyle Furge <kyle.furge@vai.org> and Karl Dykema <karl.dykema@vai.org>

References

??

See Also

writeSmc

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
str(sample)
sample[[1]]@reference <- "fileName"

## Not run:
writeSmc(sample[[1]])
smc <- readSmc("fileName-0.txt")

## End(Not run)
```

---

scanSmc *Scan through smc objects*

Description

This function scans through smc objects and returns those with specified attributes.

Usage

```
scanSmc(smcList, scanSlot = "private", scanFor = "no")
```

Arguments

smcList	list of "smc" objects
scanSlot	character - which smc slot to investigate
scanFor	character - what character string to look for

Value

a list of "smc" objects with the desired attribute

Author(s)

Karl Dykema <karl.dykema@vai.org>

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
sample[1:2] <- editSmc(sample[1:2],"creator","Joe Smith")

scanned <- scanSmc(sample,"creator","Joe Smith")
```

```
str(scanned)
```

---

smcPlot   *Plot PGSEA results*

Description

This basic function will plot results from PGSEA with easy altering of margins, colors, and text.

Usage

```
smcPlot(m, ff = NULL, skip = "NO", scale = c(-3, 3), na.color = par("bg"), margins = NULL, r.cex = NULL
```

Arguments

m	matrix - your results from PGSEA (or any other numeric matrix of data)
ff	factor - this factor corresponds to the subtypes of your samples and will control the column names
skip	character - which subtype(s) to skip from "ff"
scale	vector, length 2 - this vector sets the minimum and maximum values for the graph scale (at bottom of plot)
na.color	character - color to display in the result of an NA
margins	vector, length 4 - this vector gives the expansion values for the margins
r.cex	numeric - number giving the amount by which row names should be scaled relative to the default
c.cex	numeric - number giving the amount by which column names should be scaled relative to the default
show.grid	boolean - show grid outlines within plot?
cnames	boolean or character - vector of alternative column names
rnames	boolean or character - vector of alternative row names
grid.lty	numeric - line type of the grid lines
clust	boolean - want to cluster?
...	additional graphical parameters passed along to the plotting function

Author(s)

Karl Dykema <karl.dykema@vai.org>

Examples

```
library(PGSEA)
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
data(nbEset)

pg <- PGSEA(nbEset,cl=sample,ref=1:5)
sub <- factor(c(rep(NA,5),rep("NeuroB",5),rep("NeuroB_MYC+",5)))

smcPlot(pg,sub,scale=c(-10,10),col=.rwb,margins=c(1,1,8,13))
```

VAIgsc   *Molecular Concepts (Gene Sets) prepared at VAI*

Description

A few gene sets compiled at VAI. We have found useful in our analysis.

Usage

```
data(VAIgsc)
```

Format

The format is: chr "VAIgsc"

Source

Various sources... See individual objects for PMID, GEO accession, etc..

Examples

```
data(VAIgsc)
summary(VAIgsc)
details(VAIgsc[[1]])
```

VAImcs *Molecular Concepts prepared at VAI*

Description

A few gene sets compiled at VAI. We have found useful in our analysis.

Usage

```
data(VAImcs)
```

Format

a list of "smc" objects

Source

See individual concepts for PMID or other source information.

Examples

```
data(VAImcs)
str(VAImcs)
```

---

writeGmt *writeGmt*

Description

This function writes out SMC objects into .gmt file format

Usage

```
writeGmt(fname, cl)
```

Arguments

fname	name of the file to be written out
cl	list of SMC objects

Details

The .gmt file format is a tab delimited file format used to store gene lists. These gene lists are stored row by row. The first column is the gene set name. The second column is a brief description, and every entry after that is a gene within that gene set.

Author(s)

Kyle Furge <kyle.furge@vai.org> and Karl Dykema <karl.dykema@vai.org>

References

http://www.broad.mit.edu/gsea/doc/data_formats.html#gmt

See Also

readGmt

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
str(sample)

## Not run:
writeGmt(paste(datadir,"/output.gmt",sep=""),sample)

## End(Not run)
```

---

writeSmc   *writeSmc*

Description

This function will write out SMC objects to individual text files

Usage

```
writeSmc(x)
```

Arguments

x   an object of class SMC

Details

The file name is determined by the reference slot of the SMC object.

Author(s)

Kyle Furge <kyle.furge@vai.org> and Karl Dykema <karl.dykema@vai.org>

See Also

writeSmc

Examples

```
datadir <- system.file("extdata", package = "PGSEA")
sample <- readGmt(file.path(datadir, "sample.gmt"))
str(sample)
sample[[1]]@reference <- "fileName"

## Not run:
writeSmc(sample[[1]])

## End(Not run)
```

Index