# Package 'SNAGEE'

May 8, 2025

Version 1.48.0

Date 2013-07-16

Title Signal-to-Noise applied to Gene Expression Experiments

Author David Venet <davenet@ulb.ac.be>

Maintainer David Venet <davenet@ulb.ac.be>

Depends R (>= 2.6.0), SNAGEEdata

Suggests ALL, hgu95av2.db

Enhances parallel

Description Signal-to-Noise applied to Gene Expression Experiments.

Signal-to-noise ratios can be used as a proxy for quality of gene expression studies and samples. The SNRs can be calculated on any gene expression data set as long as gene IDs are available, no access to the raw data files is necessary. This allows to flag problematic studies and samples in any public data set.

License Artistic-2.0

biocViews Microarray, OneChannel, TwoChannel, QualityControl

URL http://bioconductor.org/

git\_url https://git.bioconductor.org/packages/SNAGEE

git\_branch RELEASE\_3\_21

git\_last\_commit ac5a58d

git\_last\_commit\_date 2025-04-15

Repository Bioconductor 3.21

Date/Publication 2025-05-07

# Contents

SNAGEE-package							 									 		2
qualSample							 									 		3
qualStudy						•	 									 		4
toSnageeFormat .							 									 		5
-																		

6

Index

SNAGEE-package

# Description

Signal-to-Noise Applied to Gene Expression Experiments

#### Details

Package:	SNAGEE
Version:	0.99.0
Date:	2012-01-26
Depends:	R (>= 2.6.0)
Imports:	SNAGEEdata
Suggests:	ALL
Enhances:	parallel
License:	Artistic-2.0
URL:	http://fleming.ulb.ac.be/SNAGEE

Index:

qualStudy	Quality of a study
qualSample	Quality of samples in a study
toSnageeFormat	Turns an Eset to a list usable by $\ensuremath{SNAGEE}$

#### Author(s)

David Venet <davenet@ulb.ac.be>

Maintainer: David Venet <davenet@ulb.ac.be>

```
# Get the list of genes
geneList = getCC()$g;
# Create a random data set
d=list(genes=geneList, data=matrix(rnorm(length(geneList)*50),ncol=50));
# Calculate its quality (it's going to be very close to 0)
qualStudy(d, disattenuate=FALSE);
# Calcuate individual sample qualities
qs = qualSample(d);
```

qualSample

#### Description

Calculate the relative quality of all samples from a study.

#### Usage

qualSample(data,mode="complete",cc=NULL,multicore=FALSE)

#### Arguments

data	The study data. If an Eset, toSnageeFormat is called on it. Otherwise, must be a list with fields 'genes' containing the vector of gene IDs (from Entrez) and 'data' containing the gene expression data.
mode	Which gene-gene correlation matrix should be used. Can be 'complete' (using all platforms) or 'woAffy' (without the Affy platforms).
сс	Can be used if wishing to use a custom gene-gene correlation matrix. Must be a list with fields 'g' containing the gene IDs and 'cc' containing the (upper triangular part of the) correlations.
multicore	Should the parallel version be used? This is based on the parallel package, if that package cannot be loaded it will fall back on single core, with a warning.

#### Details

The function calculates the quality of all samples in a study. Lower values are of lower quality. The numerical values of the study (the 'data' field) should be in log-scale, and normalized. It is recommended to used medpolish on the data.

Each gene should only appear once in the gene list. Duplicated genes must be merged before using the function. Non-finite values should also be removed first (using the impute package for instance).

#### See Also

SNAGEE, qualStudy, toSnageeFormat

```
# Get the list of genes
geneList = getCC()$g;
# Create a random data set
d=list(genes=geneList, data=matrix(rnorm(length(geneList)*50),ncol=50));
# And calculate the quality of the samples (they are all about the same)
qualSample(d);
```

qualStudy

## Description

Calculate the quality of a study.

#### Usage

qualStudy(d,mode="complete",cc=NULL,disattenuate=TRUE)

#### Arguments

d	The study data. If an Eset, toSnageeFormat is called on it. Otherwise, must be a list with fields 'genes' containing the vector of gene IDs (from NCBI's Gene DB) and 'data' containing the actual data.
mode	Which gene-gene correlation matrix should be used. Can be 'complete' (using all platforms) or 'woAffy' (without the Affy platforms).
сс	Can be used if wishing to use a custom gene-gene correlation matrix. Must be a list with fields 'g' containing the gene IDs and 'cc' containing the (upper triangular part of the) correlations.
disattenuate	Should the qualities be disattenuated?

#### Details

The function calculates the quality of a study. The numerical values of the study (the 'data' field) should be in log-scale, and normalized. It is recommended to used medpolish on the data.

Each gene should only appear once in the gene list. Duplicated genes must be merged before using the function.

The mode 'woAffy' may be useful to compare Affymetrix to not Affymetrix studies. As the median gene correlation matrix was calculated with a majority of Affymetrix platforms, those platforms tend to be given higher quality than the others with the 'complete' mode, which may be misleading.

#### See Also

SNAGEE, qualSample, linktoSnageeFormat

```
# Get the list of genes
geneList = getCC()$g;
# Create a random data set
d=list(genes=geneList, data=matrix(rnorm(length(geneList)*50),ncol=50));
# And calculate its quality (it's going to be close to 0)
qualStudy(d, disattenuate=FALSE);
```

toSnageeFormat

#### Description

Turns an Eset into a list usable by SNAGEE.

#### Usage

toSnageeFormat(data)

#### Arguments

data

An Eset. If already a list, leaves it as it is.

#### Details

The function turns an Eset into a list usable by SNAGEE. Gene ID annotations are found using the annotation slot of the Eset, and the related annotation DB. If no annotation DB can be found, gives an error.

In addition, features with identical gene IDs are averaged, and the data are medpolished.

#### See Also

SNAGEE, qualStudy, qualSample

```
# Get the list of genes
geneList = getCC()$g;
# Create a random data set
d=list(genes=geneList, data=matrix(rnorm(length(geneList)*50),ncol=50));
# And calculate its quality (it's going to be close to 0)
qualStudy(d, disattenuate=FALSE);
```

# Index

qualSample, 3, 4, 5 qualStudy, 3, 4, 5

 $\begin{array}{l} \text{SNAGEE, } 3-5 \\ \text{SNAGEE (SNAGEE-package), } 2 \\ \text{SNAGEE-package, } 2 \end{array}$ 

toSnageeFormat, 3, 4, 5