# Package 'Rfastp'

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```
Type Package
```

**Title** An Ultra-Fast and All-in-One Fastq Preprocessor (Quality Control, Adapter, low quality and polyX trimming) and UMI Sequence Parsing).

**Version** 1.16.0

**Description** Rfastp is an R wrapper of fastp developed in c++.

fastp performs quality control for fastq files. including low quality bases trimming, polyX trimming, adapter auto-detection and trimming, paired-end reads merging, UMI sequence/id handling. Rfastp can concatenate multiple files into one file (like shell command cat) and accept multiple files as input.

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**Encoding** UTF-8

LazyData true

RoxygenNote 7.1.1

biocViews QualityControl, Sequencing, Preprocessing, Software

SystemRequirements GNU make

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Suggests BiocStyle, testthat, knitr, rmarkdown

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Concatenate Fastq Files.

#### **Description**

concatenate multiple fastq files into a single file.

#### Usage

```
catfastq(output, inputFiles, append = FALSE, paired = FALSE, shuffled = FALSE)
```

# Arguments

output output file name [string] a vector of input file names [vector] inputFiles append a logical indicating append the files to a file already exists. paired a logical indicating split the input files into two halves. the first half merged into read1, the second half merged into read2. shuffled a logical indicating split the input file list into two halves. The R1/R2 files are

# Value

no returns.

#### Author(s)

Wei Wang

# **Examples**

```
pe001_read1 <- system.file("extdata","splited_001_R1.fastq.gz",</pre>
     package="Rfastp")
pe002_read1 <- system.file("extdata","splited_002_R1.fastq.gz",</pre>
     package="Rfastp")
pe003_read1 <- system.file("extdata","splited_003_R1.fastq.gz",</pre>
     package="Rfastp")
pe004_read1 <- system.file("extdata","splited_004_R1.fastq.gz",</pre>
     package="Rfastp")
pe001_read2 <- system.file("extdata","splited_001_R2.fastq.gz",</pre>
     package="Rfastp")
pe002_read2 <- system.file("extdata", "splited_002_R2.fastq.gz",</pre>
```

inteleaved in the inputFiles vector.

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```
package="Rfastp")
pe003_read2 <- system.file("extdata", "splited_003_R2.fastq.gz",</pre>
     package="Rfastp")
pe004_read2 <- system.file("extdata", "splited_004_R2.fastq.gz",</pre>
     package="Rfastp")
allR1 <- c(pe001_read1, pe002_read1, pe003_read1, pe004_read1)</pre>
allR2 <- c(pe001_read2, pe002_read2, pe003_read2, pe004_read2)
allreads <- c(allR1, allR2)
allreads_shuffled <- c(pe001_read1, pe001_read2, pe002_read1, pe002_read2,
               pe003_read1, pe003_read2, pe004_read1, pe004_read2)
outputPrefix <- tempfile(tmpdir = tempdir())</pre>
# a normal concatenation for single-end libraries.
catfastq(output = paste0(outputPrefix, "_R1.fastq.gz"), inputFiles = allR1)
# a normal concatenation for paired-end libraries.
catfastq(output = paste0(outputPrefix, "merged_paired"),
    inputFiles = allreads, paired=TRUE)
# Append to exist files (paired-end)
catfastq(output=paste0(outputPrefix, "append_paired"), inputFiles=allreads,
    append=TRUE, paired=TRUE)
# Input paired-end files are shuffled.
catfastq(output=paste0(outputPrefix, "_shuffled_paired"),
    inputFiles=allreads_shuffled, paired=TRUE, shuffled=TRUE)
```

curvePlot

Plot of Base Quality and GC Content.

#### **Description**

generate a ggplot2 object of Base Quality/GC content before and after QC.

#### Usage

```
curvePlot(json, curves = "quality_curves")
```

#### **Arguments**

json the output json of function rfastq. [json]

curves plots for Base Quality("quality\_curves") or GC content("content\_curves"). de-

fault is "quality\_curves"

#### Value

a ggplot2 object.

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#### Author(s)

Wei Wang

#### **Examples**

qcSummary

Summary of Fasta Quality Control

#### **Description**

generate a data frame of the Fastq QC summary.

# Usage

```
qcSummary(json)
```

#### **Arguments**

json

the output json of function rfastq. [json]

#### Value

a data frame.

#### Author(s)

Wei Wang

# **Examples**

rfastp

R wrap of fastp

#### **Description**

Quality control (Cut adapter, low quality trimming, polyX trimming, UMI handling, and etc.) of fastq files.

# Usage

```
rfastp(
 read1,
 read2 = "",
 outputFastq,
 unpaired = ""
  failedOut = ""
 merge = FALSE,
 mergeOut = "",
 phred64 = FALSE,
  interleaved = FALSE,
 fixMGIid = FALSE,
 adapterTrimming = TRUE,
 adapterSequenceRead1 = "auto",
 adapterSequenceRead2 = "auto",
 adapterFasta = "",
  trimFrontRead1 = 0,
  trimTailRead1 = 0,
  trimFrontRead2 = 0,
  trimTailRead2 = 0,
 maxLengthRead1 = 0,
 maxLengthRead2 = 0,
  forceTrimPolyG = FALSE,
 disableTrimPolyG = FALSE,
 minLengthPolyG = 10,
  trimPolyX = FALSE,
 minLengthPolyX = 10,
 cutWindowSize = 4,
 cutLowQualTail = FALSE,
 cutSlideWindowRight = FALSE,
  cutLowQualFront = FALSE,
  cutMeanQual = 20,
 cutFrontWindowSize = 4,
 cutFrontMeanQual = 20,
  cutTailWindowSize = 4,
 cutTailMeanQual = 20,
 cutSlideWindowSize = 4,
  cutSlideWindowQual = 20,
 qualityFiltering = TRUE,
 qualityFilterPhred = 15,
 qualityFilterPercent = 40,
 maxNfilter = 5,
```

```
averageQualFilter = 0,
  lengthFiltering = TRUE,
 minReadLength = 15,
 maxReadLength = 0,
  lowComplexityFiltering = FALSE,
 minComplexity = 30,
  index1Filter = "",
  index2Filter = "",
 maxIndexMismatch = 0,
  correctionOverlap = FALSE,
 minOverlapLength = 30,
 maxOverlapMismatch = 5,
 maxOverlapMismatchPercentage = 20,
 umi = FALSE,
  umiLoc = "",
  umiLength = 0,
  umiPrefix = "",
  umiSkipBaseLength = 0,
  umiNoConnection = FALSE,
  umiIgnoreSeqNameSpace = FALSE,
 overrepresentationAnalysis = FALSE,
 overrepresentationSampling = 20,
  splitOutput = 0,
  splitByLines = 0,
  thread = 2,
  verbose = TRUE
)
```

### **Arguments**

read1 input file name(s). [vector] read2 read2 input file name(s). [vector]

outputFastq string of /path/prefix for output fastq [string]

unpaired for PE input, output file name for reads which the mate reads failed to pass the

QC [string], default NULL, discard it. [string]

failedOut file to store reads that cannot pass the filters default NULL, discard it. [string]

merge for PE input, A logical(1) indicating whether merge each pair of reads into a

single read if they are overlaped, unmerged reads will be write to 'output' file.

Default is FALSE. the 'mergeOut' must be set if TRUE.

mergeOut under 'merge' mode, file to store the merged reads. [string]

phred64 A logical indicating whether the input is using phred64 scoring (it will be con-

verted to phred33, so the output will still be . phred33)

interleaved A logical indicating whether <read1> is an interleaved FASTQ which contains

both read1 and read2. Default is FALSE.

fixMGIid the MGI FASTQ ID format is not compatible with many BAM operation tools,

enable this option to fix it. Default is FALSE

 $adapter \\ Trimming$ 

A logical indicating whether run adapter trimming. Default is 'TRUE'

adapterSequenceRead1

the adapter for read1. For SE data, if not specified, the adapter will be auto-detected. For PE data, this is used if R1/R2 are found not overlapped.

adapterSequenceRead2

the adapter for read2 (PE data only). This is used if R1/R2 are found not over-

lapped. If not specified, it will be the same as <adapterSequenceRead1>

adapterFasta specify a FASTA file to trim both read1 and read2 (if PE) by all the sequences

in this FASTA file.

trimFrontRead1 trimming how many bases in front for read1, default is 0.

trimTailRead1 trimming how many bases in tail for read1, default is 0'

trimFrontRead2 trimming how many bases in front for read2. If it's not specified, it will follow

read1's settings

trimTailRead2 trimming how many bases in tail for read2. If it's not specified, it will follow

read1's settings

maxLengthRead1 if read1 is longer than maxLengthRead1, then trim read1 at its tail to make it as

long as maxLengthRead1 Default 0 means no limitation.

maxLengthRead2 if read2 is longer than maxLengthRead2, then trim read2 at its tail to make it as

long as maxLengthRead2. Default 0 means no limitation. If it's not specified, it

will follow read1's settings.

forceTrimPolyG A logical indicating force polyG tail trimming, trimming is only automatically

enabled for Illumina NextSeq/NovaSeq . data.

disableTrimPolyG

A logical indicating disable polyG tail trimming.

minLengthPolyG the minimum length to detect polyG in the read tail. 10 by default.

trimPolyX A logical indicating force polyX tail trimming.

minLengthPolyX the minimum length to detect polyX in the read tail. 10 by default.

cutWindowSize the window size option shared by cutLowQualFront, cutLowQualTail, or cut-

SlideWindowRight. Range: 1~1000, default: 4

cutLowQualTail A logical indiccating move a sliding window from tail (3') to front, drop the

bases in the window if its mean quality < threshold, stop otherwise. Default is

'FALSE'

cutSlideWindowRight

A logical indicating move a sliding window from front to tail, if meet one window with mean quality < threshold, drop the bases in the window and the right

part, and then stop. Default is 'FALSE'

cutLowQualFront

A logical indiccating move a sliding window from front (5') to tail, drop the bases in the window if its mean quality < threshold, stop otherwise. Default is

'FALSE'

cutMeanQual the mean quality requirement option shared by cutLowQualFront, cutLowQual-

Tail or cutSlideWindowRight. Range: 1~36, default: 20

cutFrontWindowSize

the window size option of cutLowQualFront, default to cutWindowSize if not

specified. default: 4

cutFrontMeanQual

the mean quality requirement option for cutLowQualFront, default to cutMean-

Qual if not specified. default: 20

cutTailWindowSize

the window size option of cutLowQualTail, default to cutWindowSize if not

specified. default: 4

cutTailMeanQual

the mean quality requirement option for cutLowQualTail, default to cutMean-Qual if not specified. default: 20

cutSlideWindowSize

the window size option of cutSlideWindowRight, default to cutWindowSize if not specified. default: 4

cutSlideWindowOual

the mean quality requirement option for cutSlideWindowRight, default to cut-MeanQual if not specified. default: 20

qualityFiltering

A logical indicating run quality filtering. Default is 'TRUE'.

qualityFilterPhred

the minimum quality value that a base is qualified. Default 15 means phred quality >=Q15 is qualified.

qualityFilterPercent

Maximum percents of bases are allowed to be unqualified (0~100). Default 40 means 40%

maxNfilter maximum number of N allowed in the sequence. read/pair is discarded if failed to pass this filter. Default is 5

averageQualFilter

if one read's average quality score < 'averageQualFilter', then this read/pair is discarded. Default 0 means no requirement.

lengthFiltering

A logical indicating whether run lenght filtering. Default: TRUE

minReadLength reads shorter than minReadLength will be discarded, default is 15.

 $\label{eq:maxReadLength} \mbox{maxReadLength will be discarded, default 0 means no limitation.}$ 

 ${\tt lowComplexityFiltering}$ 

A logical indicating whethere run low complexity filter. The complexity is defined as the percentage of base that is different from its next base (base[i] != base[i+1]). Default is 'FALSE'

minComplexity the threshold for low complexity filter (0~100). Default is 30, which means 30% complexity is required. (int [=30])

index1Filter specify a file contains a list of barcodes of index1 to be filtered out, one barcode per line.

index2Filter specify a file contains a list of barcodes of index2 to be filtered out, one barcode per line.

maxIndexMismatch

the allowed difference of index barcode for index filtering, default 0 means completely identical.

correctionOverlap

A logical indicating run base correction in overlapped regions (only for PE data), default is 'FALSE'

minOverlapLength

the minimum length to detect overlapped region of PE reads. This will affect overlap analysis based PE merge, adapter trimming and correction. 30 by default.

maxOverlapMismatch

the maximum number of mismatched bases to detect overlapped region of PE reads. This will affect overlap analysis based PE merge, adapter trimming and correction. 5 by default.

maxOverlapMismatchPercentage

the maximum percentage of mismatched bases to detect overlapped region of PE reads. This will affect overlap analysis based PE merge, adapter trimming and correction. Default 20 means 20%

umi A logical indicating whethere preprocessing unique molecular identifier (UMI).

Default: 'FALSE'

umiLoc specify the location of UMI, can be (index1/index2/read1/read2/per\_index/per\_read)

umiLength length of UMI if the UMI is in read1/read2.

umiPrefix an string indication the following string is UMI (i.e. prefix=UMI, UMI=AATTCG,

final=UMIAATTCG). Only letters, numbers, and '#" allowed. No prefix by de-

fault

umiSkipBaseLength

if the UMI is in read1/read2, skip 'umiSkipBaseLength' bases following UMI,

umiNoConnection

an logical indicating remove "\_" between the UMI prefix string and the UMI string. Default is FALSE.

umiIgnoreSeqNameSpace

an logical indicating ignore the space in the sequence name. Default is FALSE, the umi tag will be inserted into the sequence name before the first SPACE.

 ${\it overrepresentation} {\it Analysis}$ 

A logical indicating overrepresentation analysis. Default is 'FALSE'

overrepresentationSampling

one in 'overrepresentationSampling' reads will be computed for overrepresentation analysis (1~10000), smaller is slower, default is 20.

splitOutput number of files to be splitted (2~999). a sequential number prefix will be added

to output name. Default is 0 (no split)

splitByLines split output by limiting lines of each file(>=1000), a sequential number prefix

will be added to output name ( 0001.out.fq, 0002.out.fq...), default is 0 (dis-

abled).

thread owrker thread number, default is 2 verbose output verbose log information

# Value

returns a json object of the report.

## Author(s)

Thomas Carroll, Wei Wang

# **Examples**

```
# preprare for the input and output files.
```

 $\ensuremath{\text{\#}}$  if the output file exists, it will be OVERWRITEN.

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```
se_read1 <- system.file("extdata","Fox3_Std_small.fq.gz",package="Rfastp")
pe_read1 <- system.file("extdata","reads1.fastq.gz",package="Rfastp")
pe_read2 <- system.file("extdata","reads2.fastq.gz",package="Rfastp")</pre>
outputPrefix <- tempfile(tmpdir = tempdir())</pre>
# a normal single-end file
se_json_report <- rfastp(read1 = se_read1,</pre>
     outputFastq=paste0(outputPrefix, "_se"), thread = 4)
# merge paired-end data by overlap:
pe_json_report <- rfastp(read1 = pe_read1, read2 = pe_read2, merge = TRUE,</pre>
     outputFastq = paste0(outputPrefix, '_unpaired'),
     mergeOut = paste0(outputPrefix, '_merged.fastq.gz'))
# a clipr example
clipr_json_report <- rfastp(read1 = se_read1,</pre>
    outputFastq = paste0(outputPrefix, '_clipr'),
    disableTrimPolyG = TRUE,
   cutLowQualFront = TRUE,
    cutFrontWindowSize = 29,
    cutFrontMeanQual = 20,
    cutLowQualTail = TRUE,
    cutTailWindowSize = 1,
   cutTailMeanQual = 5,
   minReadLength = 29,
    adapterSequenceRead1 = 'GTGTCAGTCACTTCCAGCGG'
)
```

trimSummary

Summary of Fastq adapter and low quality trimming

#### **Description**

generate a data frame of the Fastq trim summary.

#### Usage

```
trimSummary(json)
```

## **Arguments**

json

the output json of function rfastq. [json]

### Value

a data frame.

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# Author(s)

Wei Wang

# **Examples**

```
outputPrefix <- tempfile(tmpdir = tempdir())
se_read1 <- system.file("extdata","Fox3_Std_small.fq.gz",package="Rfastp")
se_json_report <- rfastp(read1 = se_read1, outputFastq = outputPrefix,
    thread = 4, adapterSequenceRead1 = 'GTGTCAGTCACTTCCAGCGG')
trim_summary <- trimSummary(se_json_report)</pre>
```

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