# Package 'ASEB'

# October 17, 2024

| 00000117, 2024   |
|--|
| Title Predict Acetylated Lysine Sites  |
| <b>Version</b> 1.49.0  |
| Author Likun Wang <pre><wanglk@hsc.pku.edu.cn></wanglk@hsc.pku.edu.cn></pre> and Tingting Li <li>tt@hsc.pku.edu.cn&gt;.</li> |
| <b>Description</b> ASEB is an R package to predict lysine sites that can be acetylated by a specific KAT-family.             |
| Maintainer Likun Wang <wanglk@hsc.pku.edu.cn></wanglk@hsc.pku.edu.cn>  |
| <b>Depends</b> R ( $>= 2.8.0$ ), methods   |
| Imports graphics, methods, utils   |
| License GPL (>= 3)   |
| biocViews Proteomics   |
| Collate AllClasses.R AllGenerics.R SequenceInfo.R readSequence.R asebSites.R asebProteins.R drawStat.R drawEScurve.R         |
| LazyLoad yes   |
| git_url https://git.bioconductor.org/packages/ASEB   |
| git_branch devel   |
| git_last_commit cdcd402  |
| git_last_commit_date 2024-04-30  |
| Repository Bioconductor 3.20   |
| Date/Publication 2024-10-17  |
|  |
| Contents   |
| asebProteins asebSites drawEScurve drawStat readSequence SequenceInfo  |
| Index  |

2 asebProteins

asebProteins

prediction of all lysine sites on a specific protein that can be acetylated

#### **Description**

This function is used to predict all lysine sites on a specific protein that can be acetylated by a specific KAT-family.

# Usage

```
asebProteins(backgroundSites, prodefinedSites, testProteins, outputFile=NULL, permutationTimes=10
## S4 method for signature 'character, character, character'
asebProteins(backgroundSites, prodefinedSites, testProteins, outputFile=NULL, permutationTimes=10
## S4 method for signature 'SequenceInfo, SequenceInfo, SequenceInfo'
asebProteins(backgroundSites, prodefinedSites, testProteins, outputFile=NULL, permutationTimes=10
```

# **Arguments**

backgroundSites

SequenceInfo object or file name (character(1)) for background peptides set.

prodefinedSites

SequenceInfo object or file name (character(1)) for KAT special peptides set.

testProteins SequenceInfo object or file name (character(1)) for query Proteins set.

outputFile file name for output (character(1)).

permutation Times

permutation times (integer(1)), default and recommended: 10000.

# **Details**

This function is used to predict lysine sites that can be acetylated by a specific KAT-family. The whole process is similar with the GSEA method (permuting gene sets). Please see the references for details.

The first three arguments of method asebProteins can be SequenceInfo objects or file names. If these arguments are SequenceInfo objects, this method returns a list to the users besides an output file. Otherwise, this method processes the FASTA format files directly and outputs all results to a file. In this case, this method can process huge number of sites each time without loading any sequences to R workspace.

#### Value

The output file contains enrichment scores and P-values for each query site. The asebProteins, SequenceInfo, SequenceInfo, SequenceInfo, SequenceInfo.

results contains enrichment scores and P-values for each query site.

curveInfo contains information for enrichment score curves.

asebSites 3

#### Note

The acetylated lysine sites and their surrounding amino acids (8 on each side) are treated as acetylated peptides.

Example for peptides sequence: "KEHDDIFDKLKEAVKEE".

All input file should follow FASTA format.

#### References

Subramanian, A. et al. (2005) Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A*, **102**, 15545-15550.

Mootha, V.K. et al. (2003) PGC-1alpha-responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat Genet*, **34**, 267-273.

Guttman, M. et al. (2009) Chromatin signature reveals over a thousand highly conserved large non-coding RNAs in mammals. *Nature*, **458**, 223-227.

Li, T.T. et al. Characterization and prediction of lysine (K)-acetyl-transferase (KAT) specific acetylation sites. *Mol Cell Proteomics*, in press.

#### See Also

SequenceInfo, readSequence, asebSites, drawStat, drawEScurve.

# **Examples**

```
backgroundSites <- readSequence(system.file("extdata", "background_sites.fa", package="ASEB"))
prodefinedSites <- readSequence(system.file("extdata", "predefined_sites.fa", package="ASEB"))
testProteins <- readSequence(system.file("extdata", "proteins_to_test.fa", package="ASEB"))
resultList <- asebProteins(backgroundSites, prodefinedSites, testProteins, permutationTimes=100)
resultList$results[1:2,]</pre>
```

asebSites

prediction of lysine sites that can be acetylated

# **Description**

This function is used to predict lysine sites that can be acetylated by a specific KAT-family.

#### Usage

```
asebSites(backgroundSites, prodefinedSites, testSites, outputFile=NULL, permutationTimes=10000)
## S4 method for signature 'character, character, character'
asebSites(backgroundSites, prodefinedSites, testSites, outputFile=NULL, permutationTimes=10000)
## S4 method for signature 'SequenceInfo, SequenceInfo, SequenceInfo'
asebSites(backgroundSites, prodefinedSites, testSites, outputFile=NULL, permutationTimes=10000)
```

# Arguments

backgroundSites

SequenceInfo object or file name (character(1)) for background peptides set.

prodefinedSites

SequenceInfo object or file name (character(1)) for KAT special peptides set.

4 asebSites

testSites SequenceInfo object or file name (character(1)) for query peptides set.

outputFile file name for output (character(1)).

permutationTimes

permutation times (integer(1)), default and recommended: 10000.

#### **Details**

This function is used to predict lysine sites that can be acetylated by a specific KAT-family. It will give an enrichment score and a P-value for each query lysine site. The whole process is similar with the GSEA method (permuting gene sets). Please see the references for details.

The first three arguments of method asebSites can be SequenceInfo objects or file names. If these arguments are SequenceInfo objects, this method returns a list to the users besides an output file. Otherwise, this method processes the FASTA format files directly and outputs all results to a file. In this case, this method can process huge number of sites each time without loading any sequences to R workspace.

# Value

The output file contains enrichment scores and P-values for each query site. The asebSites, SequenceInfo, SequenceInfo also returns a list contains two data. frame objects: results and curveInfo.

results contains enrichment scores and P-values for each query site.

curveInfo contains information for enrichment score curves.

#### Note

The acetylated lysine sites and their surrounding amino acids (8 on each side) are treated as acetylated peptides.

Example for peptides sequence: "KEHDDIFDKLKEAVKEE".

All input file should follow FASTA format.

#### References

Subramanian, A. et al. (2005) Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A*, **102**, 15545-15550.

Mootha, V.K. et al. (2003) PGC-1alpha-responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat Genet*, **34**, 267-273.

Guttman, M. et al. (2009) Chromatin signature reveals over a thousand highly conserved large non-coding RNAs in mammals. *Nature*, **458**, 223-227.

Li, T.T. et al. Characterization and prediction of lysine (K)-acetyl-transferase (KAT) specific acetylation sites. *Mol Cell Proteomics*, in press.

# See Also

SequenceInfo, readSequence, asebProteins, drawStat, drawEScurve.

drawEScurve 5

# **Examples**

```
backgroundSites <- readSequence(system.file("extdata", "background_sites.fa", package="ASEB"))
prodefinedSites <- readSequence(system.file("extdata", "predefined_sites.fa", package="ASEB"))
testSites <- readSequence(system.file("extdata", "sites_to_test.fa", package="ASEB"))
resultList <- asebSites(backgroundSites, prodefinedSites, testSites, permutationTimes=100)
resultList$results[1:2,]</pre>
```

drawEScurve

draw Enriment Score curves for specific sites

# **Description**

This function is used to draw Enriment Score curves for specific sites.

# Usage

```
drawEScurve(curveInfoDataFrame, sites=NULL, max_p_value=0.1, min_es=0.2, outputDir=NULL, figKind=0.4 method for signature 'data.frame'
drawEScurve(curveInfoDataFrame, sites=NULL, max_p_value=0.1, min_es=0.2, outputDir=NULL, figKind=0.4 method
```

#### **Arguments**

curveInfoDataFrame

data.frame object: contains curve information for sites, see example.

sites character vector: only draw curves for sites with ids appear in this vector.

default: draw curves for all the sites appear in curveInfoDataFrame.

max\_p\_value numeric(1), only draw curves for sites with p-value less than this value.

min\_es numeric(1), only draw curves for sites with Enriment Score more than this value.

outputDir character(1), output directory name for all the figures.

figKind character(1), fig format: "pdf" or "jpeg".

# **Details**

This function is used to draw Enrichment Score curves for specific sites. These curves show running-sum process for calculating enrichment score. The data frame object contains curve information is given by asebSites or asebProteins.

# See Also

SequenceInfo, readSequence, asebSites, asebProteins, drawStat.

# **Examples**

```
backgroundSites <- readSequence(system.file("extdata", "background_sites.fa", package="ASEB"))
prodefinedSites <- readSequence(system.file("extdata", "predefined_sites.fa", package="ASEB"))
testSites <- readSequence(system.file("extdata", "sites_to_test.fa", package="ASEB"))
resultList <- asebSites(backgroundSites, prodefinedSites, testSites, permutationTimes=100)
drawEScurve(resultList$curveInfo, max_p_value=0.1, min_es=0.1, outputDir=tempdir(), figKind="jpeg")
cat("see figures in output dir:", tempdir(),"\n")</pre>
```

6 drawStat

| drawStat | draw P-values and enrichment scores for all lysine sites on a specific protein |
|----------|--|
|          |  |

# **Description**

This function is used to show P-values and enrichment scores for all lysine sites on a specific protein.

# Usage

```
drawStat(curveInfoDataFrame, proteinIds=NULL, outputDir=NULL, figKind=c("pdf","jpeg"))
## S4 method for signature 'data.frame'
drawStat(curveInfoDataFrame, proteinIds=NULL, outputDir=NULL, figKind=c("pdf","jpeg"))
```

# Arguments

curveInfoDataFrame

data.frame object: contains curve information for all proteins, see example.

proteinIds character vector: only draw curves for proteins with ids appear in this vector.

default: draw curves for all the proteins appear in curveInfoDataFrame.

outputDir character(1), output directory name for all the figures.

figKind character(1), fig format: "pdf" or "jpeg".

# **Details**

This function is used to draw P-values and enrichment scores for all lysine sites on a specific protein. The X-axis shows positions of all lysine sites on a specific protein, and Y-axis shows the enrichment scores  $(0\sim1)$  and P-values  $(0\sim1)$  for each lysine site. The data frame object contains curve information is given by asebProteins.

#### See Also

SequenceInfo, readSequence, asebSites, asebProteins, drawEScurve.

# **Examples**

```
backgroundSites <- readSequence(system.file("extdata", "background_sites.fa", package="ASEB"))
prodefinedSites <- readSequence(system.file("extdata", "predefined_sites.fa", package="ASEB"))
testProteins <- readSequence(system.file("extdata", "proteins_to_test.fa", package="ASEB"))
resultList <- asebProteins(backgroundSites, prodefinedSites, testProteins, permutationTimes=100)
#drawEScurve(resultList$curveInfo, max_p_value=0.5, min_es=0, outputDir=tempdir(), figKind="jpeg")
drawStat(resultList$curveInfo, outputDir=tempdir(), figKind="jpeg");
cat("see figures in output dir:", tempdir(),"\n")</pre>
```

readSequence 7

readSequence

read sequences from file

# Description

This function is used to read sequences from FASTA format file.

# Usage

```
readSequence(file)
```

# **Arguments**

file

character(1), file name for input.

The input file should follow FASTA format.

# **Details**

This function return an object of SequenceInfo that contains sequences and identifiers from FASTA format input file.

# Value

A SequenceInfo object containing sequences and identifiers from input file.

# See Also

```
{\tt SequenceInfo, asebSites, asebProteins, drawStat, drawEScurve.}
```

# **Examples**

```
ff <- system.file("extdata", "background_sites.fa", package="ASEB")
readSequence(ff)</pre>
```

SequenceInfo

"SequenceInfo" class

# **Description**

This class is used to store sequences and identifiers for lysine sites or proteins.

# **Objects from the Class**

Objects from this class are created by constructor SequenceInfo, as outlined below.

# **Slots**

```
sequences: "character" containing sequences. id: "character" containing identifiers.
```

8 SequenceInfo

# Methods

Constructor:

**SequenceInfo** signature(sequences = "character", id = "character"): Create a SequenceInfo object from sequences and their identifiers. The length of id must match that of sequences.

# See Also

read Sequence, as ebSites, as ebProteins, drawStat, drawEScurve.

# **Examples**

showClass("SequenceInfo")

# **Index**

```
* classes
    SequenceInfo, 7
* methods
    asebProteins, 2
    asebSites, 3
    drawEScurve, 5
    drawStat, 6
    readSequence, 7
asebProteins, 2, 4-8
asebProteins, character, character, character-method
        (asebProteins), 2
aseb \verb|Proteins|, SequenceInfo|, SequenceInfo|, SequenceInfo-method
        (asebProteins), 2
asebSites, 3, 3, 5-8
aseb Sites, character, character, character-method\\
        (asebSites), 3
asebSites, SequenceInfo, SequenceInfo, SequenceInfo-method
        (asebSites), 3
drawEScurve, 3, 4, 5, 6–8
drawEScurve,data.frame-method
        (drawEScurve), 5
drawStat, 3-5, 6, 7, 8
drawStat, data.frame-method (drawStat), 6
readSequence, 3-6, 7, 8
SequenceInfo, 2-7, 7
```