

# Package ‘PoDCall’

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**Type** Package

**Title** Positive Droplet Calling for DNA Methylation Droplet Digital PCR

**Version** 1.12.0

**Description** Reads files exported from 'QX Manager or QuantaSoft' containing amplitude values from a run of ddPCR (96 well plate) and robustly sets thresholds to determine positive droplets for each channel of each individual well.

Concentration and normalized concentration in addition to other metrics is then calculated for each well. Results are returned as a table, optionally written to file, as well as optional plots (scatterplot and histogram) for both channels per well written to file. The package includes a shiny application which provides an interactive and user-friendly interface to the full functionality of PoDCall.

**License** GPL-3

**Encoding** UTF-8

**RoxygenNote** 7.1.2

**Depends** R (>= 4.4)

**Imports** ggplot2, gridExtra, mclust, diptest, rlist, shiny, DT,  
LaplacesDemon, purrr, shinyjs, readr, grDevices, stats, utils

**Suggests** knitr, rmarkdown, testthat, BiocStyle

**VignetteBuilder** knitr

**biocViews** Classification, Epigenetics, ddPCR, DifferentialMethylation,  
CpGIsland, DNAMethylation,

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<code>importAmplitudeData</code>	<i>importAmplitudeData</i>
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## Description

`importAmplitudeData`

## Usage

```
importAmplitudeData(dataDirectory, skipLines = c(0, 4))
```

## Arguments

<code>dataDirectory</code>	Path to directory containing Quantasoft amplitude files from one 96 well plate. Since well coordinates are used as identifiers, files in this directory should all be from the same 96 well plate. Furthermore, there should be no other files than the amplitude files from a well plate in the directory.
<code>skipLines</code>	Number of lines to skip in amplitude data files. Must be 0 or 4 depending on software used to export data. 0 for QuantaSoft, 4 for QXmanager.

## Value

The function returns a list of dataframes named with the well ID and contains the amplitude values from the corresponding well.



podcallChannelPlot     *podcallChannelPlot*

---

### Description

Function that calls podcallScatterplot and podcallHistogram and draws a plot with both scatter plot and histogram.

### Usage

```
podcallChannelPlot(channelData, thr, channel, plotId = NULL)
```

### Arguments

channelData	Amplitude values from one channel of a well.
thr	The threshold set for channel of a well.
channel	The channel the amplitude values belong to.
plotId	A character string with title for the plot

### Value

A gtable with scatterplot and histogram

### Examples

```
## Get path to data
path <- system.file("extdata", "Amplitudes/", package="PoDCall")

## Read in data
data <- importAmplitudeData(path, skipLines=0)
data("thrTable")

## Get name of first list element and use as well ID
well_id <- names(data)[1]

## Set channel to plot
channel <- 1

## Get threshold for well_id and channel 1 (see ?thrTable)
thr <- thrTable[well_id, "thr_target"]

podcallChannelPlot(channelData=data[[well_id]][[channel]], thr, channel)
```

podcallDdpcr

*Positive Droplet Calling for ddPCR***Description**

Wrapper function that provide a complete workflow for the functionality of PoDCall. It takes path to amplitude files and sample sheet (optional), and parameters for setting threshold as input. Calls functions that read in data from files, sets threshold for each channel per well, calculates concentrations and optionally makes scatter plot and histogram for each channel per well. Results are returned as a table, optionally written to file. Plots will be written to file in a results directory if argument plots is set to TRUE.

**Usage**

```
podcallDdpcr(dataDirectory,
             sampleSheetFile=NULL,
             B=200,
             Q=9,
             refwell=1,
             ch2=TRUE,
             software=c("QuantaSoft", "QX Manager")[2],
             resultsToFile=FALSE,
             plots=FALSE,
             resPath=NULL)
```

**Arguments**

dataDirectory	Path to directory containing QuantaSoft amplitude files from one 96 well plate. Since well coordinates are used as identifiers, files in this directory should all be from the same 96 well plate. Furthermore, there can be no other files than the amplitude files from a well plate in the directory.
sampleSheetFile	File (optional) containing sample information from ddPCR experiment. This file must be a comma separated file containing the following columns: Well, Sample, TargetType and Target.
B	The number of permutations used for the Likelihood Ratio Test (default=200)
Q	A parameter for calling outliers (default=9)
refwell	reference well to calculate the shift in baseline (default=1)
ch2	Logical argument to denote channel 2 amplitudes (default=TRUE)
software	The software data was exported from, either QuntaSoft or QXmanager. Needs to be specified to ensure correct reading of data and sample sheet due to difference in formatting. (default="QX Manager")
resultsToFile	Should results be written to file(.csv)? (default=FALSE)
plots	Should plots be created and written to file? (default=FALSE)
resPath	Optional argument to provide results directory path (default=NULL)

**Value**

The function returns a table (data frame) with thresholds, droplet counts, concentration and normalized concentration. The table is optionally written to a .csv-file and plots for both channels per well can be written to files.

**Examples**

```
## Paths to data and sample sheet
dataPath <- system.file("extdata", "Amplitudes/", package="PoDCall")
ssPath <- system.file("extdata", "Sample_names.csv", package="PoDCall")

## Run PodCall
podcallResults <- podcallDdpcr(dataDirectory=dataPath,
                               sampleSheetFile=ssPath,
                               B=100, software="QuantaSoft")
```

---

podcallHistogram      *podcallHistogram*

---

**Description**

Function that make a histogram of amplitude values from one channel of a well with threshold indicated by a vertical line.

**Usage**

```
podcallHistogram(channelData, thr, channel, plotId = NULL)
```

**Arguments**

channelData	Amplitude values from one channel of a well.
thr	The threshold set for channel of a well.
channel	The channel the amplitude values belong to.
plotId	A character string with title for the plot

**Value**

A histogram of amplitude values from a channel from a well with a line indicating the set threshold.

**Examples**

```
# Get path to data
path <- system.file("extdata", "Amplitudes/", package="PoDCall")

# Read in data
data <- importAmplitudeData(path, skipLines=0)
data("thrTable")
```



```
channel=1)
```

---

```
podcallScatterplot    podcallScatterplot
```

---

### Description

Function that make a scatterplot of amplitude values from one channel of a well with threshold indicated by a horizontal line.

### Usage

```
podcallScatterplot(channelData, thr, channel, plotId = NULL)
```

### Arguments

channelData	Amplitude values from one channel of a well.
thr	The threshold set for channel of a well.
channel	The channel the amplitude values belong to.
plotId	A character string with title for the plot

### Value

A scatterplot of all droplets from a channel from a well with a line indicating the set threshold.

### Examples

```
# Get path to data
path <- system.file("extdata", "Amplitudes/", package="PoDCall")

# Read in data
data <- importAmplitudeData(path, skipLines=0)
data("thrTable")

# Get name of first list element and use as well ID
well_id <- names(data)[1]

# Set channel to plot
channel <- 1

# Get threshold for well_id and channel 1 (see ?thrTable)
thr <- thrTable[well_id, "thr_target"]

scatterplot <- podcallScatterplot(channelData=data[[well_id]][[channel]],
                                thr,
                                channel)
```



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podcallShiny      *PoDCall shiny launcher*

---

**Description**

This function launches the PoDCall shiny app in a web browser

**Usage**

```
podcallShiny()
```

**Value**

Does not return anything, but launches PoDCall shiny app

**Examples**

```
## Not run:
podcallShiny()

## End(Not run)
```

---

podcallThresholds      *podcallThresholds*

---

**Description**

Function sets threshold per channel per well and calculates concentrations. Results are returned as a data frame.

**Usage**

```
podcallThresholds(plateData,
                  nchannels=c(1,2)[2],
                  B=200,
                  Q=9,
                  refWell=1,
                  updateProgress=NULL)
```

**Arguments**

plateData	List of data frames with amplitude data from a 96 well plate
nchannels	Number of channels used in the experiment (default=2)
B	Number of permutations for the Likelihood Ratio Test (LRT) (default=200)
Q	Parameter for outlier calling (default=9)
refWell	reference well to calculate the shift in baseline (default=1)
updateProgress	function to update progress bar in shiny app (default=NULL)

**Value**

A table with results and metrics, one row per well.

**Examples**

```
## Path to example data
dataPath <- system.file("extdata", "Amplitudes/", package="PoDCall")

## Read in example data
dataList <- importAmplitudeData(dataDirectory=dataPath, skipLines=0)

## Set thresholds
thresholds <- podcallThresholds(plateData=dataList,
                                B=100)
```

---

thrTable

*PoDCall Example Threshold Table*

---

**Description**

A data.frame that contains the results of running PodCall with the amplitude data files included in the package. For testing and running of examples. See vignette for more detailed description about columns.

**Usage**

```
data("thrTable")
```

**Format**

A data.frame with 13 columns, which are:

**sample\_id** Sample ID  
**thr\_target** Threshold channel 1 (target assay)  
**thr\_ctrl** Threshold channel 2 (control assay)  
**pos\_dr\_target** Positive droplets target  
**pos\_dr\_ctrl** Positive droplets control  
**tot\_droplets** Total droplets  
**c\_target** Concentration target  
**c\_ctrl** Concentration control  
**c\_norm\_4Plex** Normalized concentration based on 4Plex control  
**c\_norm\_sg** Normalized concentration based on single gene control  
**q** Parameter Q for calling outliers  
**target\_assay** Target assay  
**ctrl\_assay** Control assay  
**ref\_well** Reference well used to set threshold

**Source**

In-house cell-line experiment.

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