Package 'CSAR'

May 17, 2024

Type Package

Title Statistical tools for the analysis of ChIP-seq data

Version 1.56.0 **Date** 2009-11-09

Author Jose M Muino

Description Statistical tools for ChIP-seq data analysis. The package includes the statistical method described in Kaufmann et al. (2009) PLoS Biology: 7(4):e1000090. Briefly, Taking the average DNA fragment size subjected to sequencing into account, the software calculates genomic single-nucleotide read-enrichment values. After normalization, sample and control are compared using a test based on the Poisson distribution. Test statistic thresholds to control the false discovery rate are obtained through random permutation.

Depends R (>= 2.15.0), S4Vectors, IRanges, GenomeInfoDb, GenomicRanges

Maintainer Jose M Muino < jose.muino@live.com>

Suggests ShortRead, Biostrings

Imports stats, utils

License Artistic-2.0

LazyLoad yes

LazyData yes

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Description

Statistical tools for ChIP-seq data analysis.

The package is oriented to plant organisms, and compatible with standard file formats in the plant research field.

Details

Package: CSAR
Type: Package
Version: 1.0
Date: 2009-11-09
License: Artistic-2.0
LazyLoad: yes

Author(s)

Jose M Muino

Maintainer: Jose M Muino <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

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Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo
nhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))
nhitsC<-mappedReads2Nhits(controlSEP3_test,file="controlSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))</pre>
##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC,sample=nhitsS)</pre>
##We calculate the candidate read-enriched regions
win<-sigWin(test)</pre>
##We generate a wig file of the results to visualize tehm in a genome browser
score2wig(test,file="test.wig")
##We calculate relative positions of read-enriched regions regarding gene position
d<-distance2Genes(win=win,gff=TAIR8_genes_test)</pre>
##We calculate table of genes with read-enriched regions, and their location
genes<-genesWithPeaks(d)</pre>
##We calculate two sets of read-enrichment scores through permutation
permutatedWinScores(nn=1,sample=sampleSEP3_test,control=controlSEP3_test,fileOutput="test",chr=c("CHR1v0121200
permutatedWinScores(nn=2,sample=sampleSEP3_test,control=controlSEP3_test,fileOutput="test",chr=c("CHR1v0121200
###Next function will get all permutated score values generated by permutatedWinScores function.
##This represent the score distribution under the null hypotesis and therefore it can be use to control the error of
nulldist<-getPermutatedWinScores(file="test",nn=1:2)</pre>
##From this distribution, several cut-off values can be calculated to control the error of our test.
##Several functions in R can be used for this purpose.
##In this package we had implemented a simple method for the control of the error based on FDR"
getThreshold(winscores=values(win)$score,permutatedScores=nulldist,FDR=.01)
```

ChIPseqScore

Calculate read-enrichment scores for each nucleotide position

Description

Calculate read-enrichment scores for each nucleotide position

Usage

```
ChIPseqScore(control, sample, backg = -1, file = NA, norm = 3 * 10^9, test = "Ratio", times=1e6, digits=2"
```

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Arguments

control data.frame structure obtained by mappedReads2Nhits data.frame structure obtained by mappedReads2Nhits sample backg Due low coverage in the control, there could be regions with no hits. Any region with a hit value lower than backg in the control will be set to the value of file Name of the file where you wan to save the results (if desired) Integer value. Number of hits will be reported by number of hits per norm nunorm cleotides test Use a score based on the poisson distribution ("Poisson") or in the ratio ("Ratio") To be memory efficient, CSAR will only upload to the RAM memory fragments times of length times. A bigger value means more RAM memory needed but whole process will be faster

Number of decimal digits used to report the score values

Details

digits

Different sequencing efforts yield different number of sequenced reads, for this reason the "number of hits" at each nucleotide position is normalized by the total number of nucleotides sequenced. Subsequently, the number of hits for the sample is normalize to have the same mean and variance than the control, for each chromosome independently or for the whole set of chromosomes (depending of the value of normEachChrInd). Due low coverage, there could be regions with no hits. Any region with a hit value lower than backg in the control will be set to the value of backg For each nucleotide position, a read-enrichment score will be calculated with the Poisson test, or with the ratio.

Value

A list to be used for other functions of the CSAR package

chr Chromosme names
chrL Chromosme length (bp)

filenames Name of the files where the score values are storaged

digits Score values storaged on the files need to be divided by 10^digits

Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

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See Also

CSAR-package

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromonhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))</pre>
```

 $nhits C < -mapped Reads 2 Nhits (control SEP3_test, file="control SEP3_test", chr=c("CHR1v01212004"), chrL=c(10000)) \\$

```
##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC,sample=nhitsS)</pre>
```

distance2Genes	Calculate relative positions of read-enriched regions regarding gene
	position

Description

Calculate relative positions of read-enrichment regions regarding gene position

Usage

```
distance2Genes(win, gff, t = 1, d1 = -3000, d2 = 1000)
```

Arguments

win	GRange structure obtained with the function sigWin
gff	Data.frame structure obtained after loading a desired gff file
t	Integer. Only distances of read-enriched regions with a score bigger than t will be considered
d1	Negative integer. Minimum relative position regarding the start of the gene to be considered
d2	Positive integer. Maximum relative position regarding the end of the gene to be considered

Value

data.frame structure where each row represents one relative position, and each column being:

peakName	read-enriched region name
p1	relative position regarding the start of the gene
p2	relative position regarding the end of the gene
gene	name of the gene
le	length (bp) of the gene

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Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

genesWithPeaks, CSAR-package

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo nhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))
nhitsC<-mappedReads2Nhits(controlSEP3_test,file="controlSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))

##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC,sample=nhitsS)

##We calculate the candidate read-enriched regions
win<-sigWin(test)

##We calculate relative positions of read-enriched regions regarding gene position
d<-distance2Genes(win=win,gff=TAIR8_genes_test)</pre>
```

 ${\tt genesWithPeaks}$

Provide table of genes with read-enriched regions, and their location

Description

Provide table of genes with read-enriched regions, and their location

Usage

```
genesWithPeaks(distances)
```

Arguments

distances

data.frame structure obtained by distances2Genes

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Details

This function report for each gene, the maximum peak score in different regions near of the gene. The input of the function is the distances between genes and peaks calculated by distance2Genes

Value

data.frame structure with each coloumn being:

name	name of the gene
max3kb1kb	maximum score value for the region 3Kb upstream to 1Kb dowstream
u3000	maximum score value for the region 3Kb upstream to 2Kb upstream
u2000	maximum score value for the region 2Kb upstream to 1Kb upstream
u1000	maximum score value for the region 1Kb upstream to 0Kb upstream
d0	maximum score value for the region 0Kb upstream to 0Kb dowstream
d1000	maximum score value for the region 0Kb dowstream to 1Kb dowstream

Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

distance2Genes,CSAR-package

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo nhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))
nhitsC<-mappedReads2Nhits(controlSEP3_test,file="controlSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))

##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC,sample=nhitsS)

##We calculate the candidate read-enriched regions
win<-sigWin(test)

##We calculate relative positions of read-enriched regions regarding gene position
d<-distance2Genes(win=win,gff=TAIR8_genes_test)</pre>
```

##We calculate table of genes with read-enriched regions, and their location
genes<-genesWithPeaks(d)</pre>

getPermutatedWinScores

Obtain the read-enrichment score distribution under the null hypothesis

Description

Obtain the read-enrichment score distribution under the null hypothesis

Usage

```
getPermutatedWinScores(file, nn)
```

Arguments

file Name of the file generated by permutatedWinScores

nn ID for the multiple permutation process

Value

Numeric vector of score values under permutation

Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

CSAR-package, permutatedWinScores

getThreshold 9

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo nhitsS<-mappedReads2Nhits(sampleSEP3_test, file="sampleSEP3_test", chr=c("CHR1v01212004"), chrL=c(10000))
nhitsC<-mappedReads2Nhits(controlSEP3_test, file="controlSEP3_test", chr=c("CHR1v01212004"), chrL=c(10000))

##We calculate two sets of read-enrichment scores through permutation
permutatedWinScores(nn=1, sample=sampleSEP3_test, control=controlSEP3_test, fileOutput="test", chr=c("CHR1v0121200 permutatedWinScores(nn=2, sample=sampleSEP3_test, control=controlSEP3_test, fileOutput="test", chr=c("CHR1v0121200 permutatedWinScores function will get all permutated score values generated by permutatedWinScores function.
##This represent the score distribution under the null hypotesis and therefore it can be use to control the error of</pre>
```

getThreshold Calculate the threshold value corresponding to control FDR at a desired level

Description

Calculate the threshold value corresponding to control FDR at a desired level

Usage

getThreshold(winscores, permutatedScores, FDR)

nulldist<-getPermutatedWinScores(file="test",nn=1:2)

Arguments

winscores Numeric vector with score values obtained from the sigWin function permutatedScores

Numeric vector with the permutated read-enrichment score values

FDR Numeric value with the desired FDR control

Details

This is a very simple function to obtain the threshold value of our test statistic controlling FDR at a desired level. Other functions implemented in R (eg: multtest) could be more sophisticated. Basically, for each possible threshold value, the proportion of error type I is calculated assuming that the permutated score distribution is a optimal estimation of the score distribution under the null hypothesis. This is, the proportion of permutated scores exceding the considered threshold value is used as an estimation of the error type I of our statisitic. FDR is obtained as the ratio of the proportion of error type I by the proportion of significant tests.

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Value

A table with the columns being:

threshold The threshold value

p-value The p-value obtained from the permutated score ditribution

FDR The FDR control obtained using threshold

Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)

See Also

CSAR-package,getPermutatedWinScores, sigWin

Examples

```
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo
nhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))</pre>
nhitsC<-mappedReads2Nhits(controlSEP3_test,file="controlSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))
##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC, sample=nhitsS)
##We calculate the candidate read-enriched regions
win<-sigWin(test)</pre>
##We calculate two sets of read-enrichment scores through permutation
permutatedWinScores(nn=1,sample=sampleSEP3_test,control=controlSEP3_test,fileOutput="test",chr=c("CHR1v0121200
permutatedWinScores(nn=2,sample=sampleSEP3_test,control=controlSEP3_test,fileOutput="test",chr=c("CHR1v0121200
###Next function will get all permutated score values generated by permutatedWinScores function.
##This represent the score distribution under the null hypotesis and therefore it can be use to control the error of
nulldist<-getPermutatedWinScores(file="test",nn=1:2)</pre>
##From this distribution, several cut-off values can be calculated to control the error of our test.
##Several functions in R can be used for this purpose.
##In this package we had implemented a simple method for the control of the error based on FDR"
getThreshold(winscores=values(win)$score,permutatedScores=nulldist,FDR=.01)
```

loadMappedReads 11

Description

This function load the output file of a read mapping software (eg:SOAP)

Usage

```
loadMappedReads(file, format = "SOAP", header = FALSE)
```

Arguments

file File name to load

format Format of the file. "SOAP" for the output of the soap software and "MAQ"

for the maq software. Other user formats can be provided as a character vector for the file column names. Columns named: "Nhits", "lengthRead", "strand",

"chr", and "pos" are needed.

header Logical value indicating if the first line of the file should be skipped (TRUE) or

not (FALSE)

Value

data.frame structure that can be used by mappedReads2Nhits

Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

CSAR-package

Examples

```
##We load the mapped reads:
#sample<-loadMappedReads(file=file,format="SOAP",w=300,header=F)
##where file is the name and path of the output file of the mapping process.</pre>
```

mappedReads2Nhits

mappedReads2Nhits	Calculate number of overlapped extended reads per nucleotide position
-------------------	-----------------------------------------------------------------------

Description

Calculate number of overlapped extended reads per nucleotide position

Usage

```
mapped Reads 2 Nhits (input, file, chr = c("chr1", "chr2", "chr3", "chr4", "chr5"), chrL = "TAIR9", w = 3000 chr2", "chr4", "chr5"), chrL = "TAIR9", w = 3000 chr2", "chr4", "chr5"), chrL = "TAIR9", w = 3000 chr2", "chr4", "chr5"), chrL = "TAIR9", w = 3000 chr2", "chr4", "chr5"), chrL = "TAIR9", w = 3000 chr2", "chr4", "chr5"), chr2", "chr5", "chr4", "chr5"), chr4", "chr5"), chr4", "chr5", "chr
```

Arguments

input	data loaded with loadMappedReads or an AlignedRead object from the Short-Read package
file	Name of the file where the results will be saved. If NA the results will not be saved in a file.
chr	Character vector containing the chromosome names as identified on input.
chrL	Numeric vector containing the length (bp) of the chromosomes. It should be in the same order than chr
W	Integer corresponding to the desired length of the extended reads. An advised value will be the average fragment length of the DNA submitted to sequence (usually 300 bp).
considerStrand	Character value.
	"Minimum"=>Default value. Report the minimum number of hits at each nucleotide position for both strands.
	"Foward"=> Report the number of hits at each nucleotide position for the "foward" strands (the one denoted as "+" in q).
	"Reverse"=>Report the number of hits at each nucleotide position for the "reverse" strands (the one denoted as "-" in q).
	"Sum"=>Report the sum of number of hits at each nucleotide position for both strands.
uniquelyMapped	Logic value, If TRUE, only consider uniquely mapped reads.
uniquePosition	Logic value. If TRUE, only consider reads mapped in different positions.

Value

A list to be used for other functions of the CSAR package

chr	Chromosme names
chrL	Chromosme length (bp)
chrL_0	Number of nucleotide positions with at least one extended read
chrL_0	Number of nucleotide positions with at least one extended read

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filenames	Name of the files where the Nhits values are storaged
c1	Sum of all the Nhits values for each chromosome
c2	Sum of all the Nhits square values for each chromosome

Author(s)

```
Jose M Muino, <jose.muino@wur.nl>
```

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

CSAR-package

Examples

```
#For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009) data("CSAR-dataset");
```

#We calculate the number of hits for each nucleotide posotion for the sample. We do that just for chromosome chr1, ar nhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))

permutatedWinScores Calculate scores for permutated read-enriched regions

Description

Calculate scores for permutated read-enriched regions

Usage

```
permutatedWinScores(nn = 1, control, sample, fileOutput, chr = c("chr1", "chr2", "chr3", "chr4", "chr5"
```

Arguments

nn ID to identify each permutation

control data.frame structure obtained by loading the mapped reads with the function

LoadMappedReads()

sample data.frame structure obtained by loading the mapped reads with the function

LoadMappedReads()

fileOutput Name of the file were the results will be written

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chr Character vector containing the chromosome names as identified on q. chrL Numeric vector containing the length (bp) of the chromosomes. It should be in the same order than chr Integer corresponding to the desired length of the extended reads. considerStrand Character value. "Minimum"=>Default value. Report the minimum number of hits at each nucleotide position for both strands. "Foward"=> Report the number of hits at each nucleotide position for the "foward" strands (the one denoted as "+" in q). "Reverse"=>Report the number of hits at each nucleotide position for the "reverse" strands (the one denoted as "-" in q). "Sum"=>Report the sum of number of hits at each nucleotide position for both strands. uniquelyMapped Logic value, If TRUE, only consider unquely mapped reads. uniquePosition Logic value. If TRUE, only consider reads mapped in different positions. Integer value. Number of hits will be reported by number of hits per norm nunorm cleotides backg Any region with a hit value lower than backg in the control will be set to the value of backg t Numeric value. Read-enriched regions are calculated as genomic regions with score values bigger than t Integer value. The maximum gap allowed between regions. Regions that are g less than g bps away will be merged. times To be memory efficient, CSAR will only upload to the RAM memory fragments of length times. A bigger value means more RAM memory needed but whole process will be faster

Number of decimal digits used to report the score values

Details

digits

test

The parameter values should be the same than the one used in sigWin, ChIPseqScore, and mappedReads2Nhits. The label "control" and "sample" is asigned to each read to identify from which group they came. Labels are randomly permutated, and read-enriched regions for this new permuated dataset are calculated.

Use a score based on the poisson distribution ("Poisson") or in the ratio ("Ratio")

Value

The file filePutput is created with its values being the permuated score values.

Author(s)

Jose M Muino, <jose.muino@wur.nl>

sampleSEP3_test 15

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

CSAR-package,getPermutatedWinScores

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
```

##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo nhitsS<-mappedReads2Nhits(sampleSEP3_test, file="sampleSEP3_test", chr=c("CHR1v01212004"), chrL=c(10000)) nhitsC<-mappedReads2Nhits(controlSEP3_test, file="controlSEP3_test", chr=c("CHR1v01212004"), chrL=c(10000))</pre>

##We calculate two sets of read-enrichment scores through permutation
permutatedWinScores(nn=1,sample=sampleSEP3_test,control=controlSEP3_test,fileOutput="test",chr=c("CHR1v0121200
permutatedWinScores(nn=2,sample=sampleSEP3_test,control=controlSEP3_test,fileOutput="test",chr=c("CHR1v0121200

sampleSEP3_test

Partial dataset of a ChIP-seq experiment

Description

Partial dataset of a Solexa DNA library obtained from a ChIP-seq experiment in Arabidopsis

Source

Kaufmann et al. (2009) Target Genes of the MADS Transcription Factor SEPALLATA3: Integration of Developmental and Hormonal Pathways in the \$Arabidopsis\$ Flower. PLoS Biol 7:e1000090

Examples

data(CSAR-dataset)

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score2wig	Save the read-enrichment scores at each nucleotide position in a .wig file format

Description

Save the read-enrichment scores at each nucleotide position in a .wig file format that can be visualize by a genome browser (eg: Integrated Genome Browser)

Usage

```
score2wig(experiment, file, t = 2, times = 1e6,description="", name="")
```

Arguments

experiment	Output of the function ChIPseqScore
file	Name of the output .wig file
t	Only nucleotide positions with a read-enrichment score bigger than t will be reported
times	To be memory efficient, CSAR will only upload to the RAM memory fragments of length times. A bigger value means more RAM memory needed but whole process will be faster $ \frac{1}{2} \left(\frac{1}{2} \right) = \frac{1}{2} \left(\frac{1}{2} \right) \left(\frac{1}{2$
description	Character. It adds a description to the wig file. The description will be shown by the genome browser used to visualize the wig file.
name	Character. It adds a wig to the wig file. The name will be shown by the genome browser used to visualize the wig file.

Value

None. Results are printed in a file

Author(s)

Jose M Muino, <jose.muino@wur.nl>

References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

CSAR-package

sigWin 17

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
##We calculate the number of hits for each nucleotide position for the control and sample. We do that just for chromo nhitsS<-mappedReads2Nhits(sampleSEP3_test,file="sampleSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))
nhitsC<-mappedReads2Nhits(controlSEP3_test,file="controlSEP3_test",chr=c("CHR1v01212004"),chrL=c(10000))
##Since we will not need the raw data anymore, we could delete it from the RAM memory
rm(sampleSEP3_test,controlSEP3_test);gc(verbose=FALSE)
##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC,sample=nhitsS)

##We generate a wig file of the results to visualize them in a genome browser
score2wig(test,file="test.wig")</pre>
```

sigWin

Calculate regions of read-enrichment

Description

Calculate regions of read-enrichment

Usage

```
sigWin(experiment, t = 1, g = 100)
```

Arguments

experiment	Output of the function ChIPseqScore
t	Numeric value. Read-enriched regions are calculated as genomic regions with score values bigger than \ensuremath{t}
g	Integer value. The maximum gap allowed between regions. Regions that are less than g bps away will be merged.

Value

An object of type'GRange' with its values being:

seqnames	Chromosome name
ranges	An IRanges object indicating start and end of the read-enriched region
posPeak	Position of the maximum score value on the read-enriched region
score	Maximum score value on the read-enriched region

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References

Muino et al. (submitted). Plant ChIP-seq Analyzer: An R package for the statistical detection of protein-bound genomic regions.

Kaufmann et al.(2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the Arabidopsis flower. PLoS Biology; 7(4):e1000090.

See Also

CSAR-package

Examples

```
##For this example we will use the a subset of the SEP3 ChIP-seq data (Kaufmann, 2009)
data("CSAR-dataset");
```

##We calculate the number of hits for each nucleotide posotion for the control and sample. We do that just for chromo nhitsS<-mappedReads2Nhits(sampleSEP3_test, file="sampleSEP3_test", chr=c("CHR1v01212004"), chrL=c(10000)) nhitsC<-mappedReads2Nhits(controlSEP3_test, file="controlSEP3_test", chr=c("CHR1v01212004"), chrL=c(10000))</pre>

##We calculate a score for each nucleotide position
test<-ChIPseqScore(control=nhitsC,sample=nhitsS)</pre>

##We calculate the candidate read-enriched regions
win<-sigWin(test)</pre>

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