

# Package ‘GGtools’

April 5, 2014

**Title** software and data for analyses in genetics of gene expression

**Version** 4.10.0

**Author** VJ Carey <stvjc@channing.harvard.edu>

**Description** software and data for analyses in genetics of gene expression and/or DNA methylation

**Suggests** GGdata, illuminaHumanv1.db, SNPlocs.Hsapiens.dbSNP.20120608

**Depends** R (>= 2.14), stats4, GGBase (>= 3.19.7), IRanges, GenomicRanges, Rsamtools

**Imports**

methods, utils, stats, BiocGenerics, snpStats, ff, AnnotationDbi, Biobase, bit, VariantAnnotation

**Enhances** MatrixEQTL

**Maintainer** VJ Carey <stvjc@channing.harvard.edu>

**License** Artistic-2.0

**biocViews** Genetics, GeneExpression, GeneticVariability, SNP

**LazyLoad** yes

**Collate** AllClasses.R AllGenerics.R eqtlTests.R managers.R topFeats.R  
gwSnpTests.R snpsCisToGenes.R relocate.R topSnps.R  
snplocsDefault.R transutils.R vcfutils.R eqtlEstimates.R  
alleq.R meta.R eqME.R meta.all.R best.trans.eQTLs.R  
meta.transScores.R summInfra.R bindmaf.R fdr.all.cis.R pifdr.R  
getFDR.R cisConfig.R thinclass.R harvest.R scoredist.R

## R topics documented:

|                  |    |
|------------------|----|
| GGtools-package  | 2  |
| All.cis          | 3  |
| b1               | 4  |
| best.cis.eQTLs   | 6  |
| best.trans.eQTLs | 10 |

|                                  |    |
|----------------------------------|----|
| bindmaf . . . . .                | 12 |
| CisConfig-class . . . . .        | 13 |
| cisRun-class . . . . .           | 15 |
| collectBest . . . . .            | 16 |
| eqtlTests . . . . .              | 18 |
| eqtlTestsManager-class . . . . . | 19 |
| ex . . . . .                     | 21 |
| getCisMap . . . . .              | 22 |
| gwSnpTests . . . . .             | 24 |
| pifdr . . . . .                  | 25 |
| richNull . . . . .               | 26 |
| sensanal . . . . .               | 27 |
| sensiCisInput-class . . . . .    | 28 |
| sensiCisOutput-class . . . . .   | 29 |
| snplocsDefault . . . . .         | 30 |
| strMultPop . . . . .             | 30 |
| transManager-class . . . . .     | 31 |
| transScores . . . . .            | 32 |
| transTab . . . . .               | 34 |
| vcf2sm . . . . .                 | 35 |

**Index** **36**

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|                 |  |
|-----------------|--|
| GGtools-package | <i>software and data for analyses in genetics of gene expression</i> |
|-----------------|--|

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**Description**

software and data for analyses in genetics of gene expression

**Details**

Package: GGtools  
Version: 4.2.26  
Suggests: GGdata, illuminaHumanv1.db  
Depends: R (>= 2.14), GGBase (>= 3.16.1)  
Imports: methods, snpStats, ff, IRanges, GenomicRanges, AnnotationDbi, Biobase, Rsamtools, bit, VariantAnnotation  
License: Artistic-2.0  
LazyLoad: yes  
Packaged: 2012-01-18 03:39:51 UTC; stvjc  
Collate: AllClasses.R AllGenerics.R eqtlTests.R managers.R topFeats.R gwSnpTests.R snpsCisToGenes.R relocate.R top  
Built: R 2.15.0; ; 2012-02-06 17:22:52 UTC; unix

Index:

best.cis.eQTLs            collect genewise best scoring eQTL

|                        |  |
|------------------------|--|
| eqtlTests              | compute association statistics between all probes and SNP in an smlSet instance                                    |
| eqtlTestsManager-class | Class "eqtlTestsManager"   |
| ex                     | ExpressionSet instance for illustrating integrative smlSet container   |
| getCisMap              | create, using Bioconductor annotation resources, a structure that enumerates SNP in the vicinity of (cis to) genes |
| gwSnpTests             | execute a series of tests for association between genotype and expression  |
| strMultPop             | serialization of a table from Strangers multipopulation eQTL report  |

The package depends on GGBase, which includes additional infrastructure for integrative data structures and data filtering.

### Author(s)

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### See Also

[getSS](#) for acquiring containers for integrative data on genetics of expression.

### Examples

```
## Not run:
# acquire chromosome 20 genotypes and all expression data for
# 90 CEU samples as published at Wellcome Trust GENEVAR and
# HapMap phase II
c20 = getSS("GGtools", "20")
# perform a focused eQTL search
t1 = gwSnpTests(genesym("CPNE1")~male, c20)
# get best hits
topSnps(t1)

## End(Not run)
```

---

|         |   |
|---------|---|
| All.cis | <i>function that computes score tests for all SNP cis to genes, with flexible filtering</i> |
|---------|---|

---

### Description

function that computes score tests for all SNP cis to genes, with flexible filtering

**Usage**

```
All.cis( config = new("CisConfig"), ... )
```

**Arguments**

```
config      instance of class CisConfig-class
...         passed to eqtlTests
```

**Details**

returns score statistics for associations of all SNP cis to genes, in a GRanges instance, with range names given by probes; metadata supplied SNP location, name, and score

**Value**

instance of [cisRun-class](#)

**Examples**

```
## Not run:
cc = new("CisConfig")
chrnames(cc) = "21"
lkp = try(library(parallel))
if (!inherits(lkp, "try-error")) {
  nc = min(10, detectCores())
  options(mc.cores=nc)
  geneApply(cc) = mclapply
}
estimates(cc) = FALSE
unix.time(f1 <- All.cis( cc ))

## End(Not run)
```

---

b1 *mcwBestCis instances, integrative analysis output containers generated by GGtools vignette*

---

**Description**

integrative analysis output containers generated by GGtools vignette

**Usage**

```
data(b1)
```

**Format**

The format is:

```

Formal class 'mcwBestCis' [package "GGtools"] with 9 slots
..@ scoregr :Formal class 'GRanges' [package "GenomicRanges"] with 6 slots
.. ..@ seqnames :Formal class 'Rle' [package "IRanges"] with 4 slots
.. .. ..@ values : Factor w/ 1 level "20": 1
.. .. ..@ lengths : int 50
.. .. ..@ elementMetadata: NULL
.. .. ..@ metadata : list()
.. ..@ ranges :Formal class 'IRanges' [package "IRanges"] with 6 slots
.. .. ..@ start : int [1:50] 24280834 61665697 352356 61679079 45286150 55187941 38766161
10871477 56570242 13304639 ...
.. .. ..@ width : int [1:50] 2090785 2005619 2021461 2001901 2129211 2007692 2038197
2035767 2012068 2013675 ...
.. .. ..@ NAMES : chr [1:50] "GI_34147330-S" "hmm26961-S" "GI_17149835-I" "GI_31077201-
S" ...
.. .. ..@ elementType : chr "integer"
.. .. ..@ elementMetadata: NULL
.. .. ..@ metadata : list()
.. ..@ strand :Formal class 'Rle' [package "IRanges"] with 4 slots
.. .. ..@ values : Factor w/ 3 levels "+","-","*": 3
.. .. ..@ lengths : int 50
.. .. ..@ elementMetadata: NULL
.. .. ..@ metadata : list()
.. ..@ elementMetadata:Formal class 'DataFrame' [package "IRanges"] with 6 slots
.. .. ..@ rownames : NULL
.. .. ..@ nrows : int 50
.. .. ..@ listData :List of 6
.. .. .. ..$ score : num [1:50] 36.5 16.8 16.7 14.8 9.8 ...
.. .. .. ..$ snpid : chr [1:50] "rs6037097" "rs3810504" "rs13043344" "rs13044229" ...
.. .. .. ..$ snploc : int [1:50] 25347221 62678549 544417 61738288 46369894 56551001
38879237 12177765 57565943 15056960 ...
.. .. .. ..$ radiusUsed: num [1:50] 1e+06 1e+06 1e+06 1e+06 1e+06 1e+06 1e+06 1e+06 1e+06
1e+06 ...
.. .. .. ..$ nsnp : int [1:50] 1387 783 2382 766 2157 2238 1658 2459 1675 1868 ...
.. .. .. ..$ fdr : num [1:50] 0 0 0 0.25 0.717 ...
.. .. ..@ elementType : chr "ANY"
.. .. ..@ elementMetadata: NULL
.. .. ..@ metadata : list()
.. ..@ seqinfo :Formal class 'Seqinfo' [package "GenomicRanges"] with 4 slots
.. .. ..@ seqnames : chr "20"
.. .. ..@ seqlengths : int NA
.. .. ..@ is_circular: logi NA
.. .. ..@ genome : chr NA
.. .. ..@ metadata : list()
..@ allperm : num [1:100] 15.8 13.9 13.8 13.2 13 ...
..@ extra : NULL
..@ chromUsed: chr "20"

```

```

..@ theCall : language best.cis.eQTLs(smpack = "GGdata", rhs = ~male, folderstem = "db2", radius
= 1e+06, chrnames = "20", geneApply = mclapply, snpannopk = snplocsDefault(), ...
..@ smFilter :function (x)
.. ..- attr(*, "srcref")=Class 'srcref' atomic [1:8] 413 8 413 50 8 50 4 4
.. ..- attr(*, "srcfile")=Classes 'srcfilecopy', 'srcfile' <environment: 0xf5c0cc0>
..@ nperm : num 2
..@ globalMap:<environment: 0xf5c0538>
..@ testCount: int 83969

```

## Details

As created in GGtools.Rnw vignette code, with sharply curtailed searches

## Examples

```

data(b1)
b1

```

---

|                |   |
|----------------|---|
| best.cis.eQTLs | <i>collect genewise best scoring eQTL</i> |
|----------------|---|

---

## Description

collect genewise best scoring eQTL

## Usage

```

best.cis.eQTLs(smpack = "GGdata", rhs = ~1,
  folderstem = "cisScratch", radius = 50000,
  shortfac = 100,
  chrnames = as.character(1:22),
  smchrpref = "", gchrpref = "", schrpref = "ch",
  geneApply = lapply, geneannopk = "illuminaHumanv1.db",
  snpannopk = snplocsDefault(),
  smFilter = function(x) nsFilter(MAFfilter(x, lower = 0.05), var.cutoff = 0.97), nperm = 2,
  useME=FALSE, excludeRadius=NULL, exFilter=function(x)x,
  keepMapCache=FALSE, getDFFITS=FALSE, SSgen = GGBase::getSS)

```

```

All.cis.eQTLs(maxfdr = 0.05, inbestcis = NULL, smpack = "GGdata",
  rhs = ~1, folderstem = "cisScratch", radius = 50000,
  shortfac = 100,
  chrnames = as.character(1:22),
  smchrpref = "", gchrpref = "", schrpref = "ch",
  geneApply = lapply, geneannopk = "illuminaHumanv1.db",
  snpannopk = snplocsDefault(),
  smFilter4cis = function(x) nsFilter(MAFfilter(clipPCs(x,

```

```

    1:10), lower = 0.05), var.cutoff = 0.85),
  smFilter4all = function(x) MAFFilter(clipPCs(x,
    1:10), lower = 0.05),
  nperm = 2, excludeRadius=NULL, exFilter=function(x)x,
  SSgen = GGBase::getSS)

meta.best.cis.eQTLs(smpackvec = c("GGdata", "hmyriB36"), rhslist = list(~1,
  ~1), folderstem = "cisScratch", radius = 50000, shortfac = 100,
  chrnames = as.character(1:22), smchrpref = "", gchrpref = "",
  schrpref = "ch", geneApply = lapply, geneannopk = "illuminaHumanv1.db",
  snpannopk = snplocsDefault(), SMFilterList = list(
  function(x) nsFilter(MAFFilter(x, lower = 0.05), var.cutoff = 0.97),
  function(x) nsFilter(MAFFilter(x, lower = 0.05), var.cutoff = 0.97) ),
  exFilterList = list(function(x)x, function(x)x),
  nperm = 2, excludeRadius=NULL)

meta.All.cis.eQTLs(minchisq, smpackvec = c("GGdata", "hmyriB36"),
  rhslist = list(~1, ~1), folderstem = "cisScratch",
  radius = 50000, shortfac=100, chrnames = as.character(1:22), smchrpref = "",
  gchrpref = "", schrpref = "ch", geneApply = lapply,
  geneannopk = "illuminaHumanv1.db",
  snpannopk = snplocsDefault(),
  SMFilterList = list(function(x) nsFilter(MAFFilter(x,
    lower = 0.05), var.cutoff = 0.97), function(x)
    nsFilter(MAFFilter(x, lower = 0.05), var.cutoff =
    0.97)),
  exFilterList = list(function(x) x, function(x)
    x),
  nperm = 2)

chromsUsed(x)

fdr(x)

fullreport(x, type, ...)

getAll(x)

getBest(x)

getCall(x)

```

### Arguments

|           |   |
|-----------|---|
| smpack    | character string naming a package to which <code>getSS</code> can be applied to extract <code>smlSet-class</code> instances |
| smpackvec | vector of character strings naming packages that can be used as smpack values   |

|                           |  |
|---------------------------|--|
|                           | in a series of <code>best.cis.eQTLs</code> calls, one per population for meta-analysis   |
| <code>rhs</code>          | R model formula, with no dependent variable, that will be used with <code>snp.rhs.tests</code> to adjust GWAS tests for each expression probe  |
| <code>rhslist</code>      | a list of model formulae to be used as <code>rhs</code> in a series of <code>best.cis.eQTLs</code> calls, one per population for meta-analysis   |
| <code>folderstem</code>   | prefix of the folder name to be used to hold <code>ff</code> archives of test results  |
| <code>radius</code>       | coding extent of each gene will be extended in both directions by <code>radius</code> bases, and only SNP within these limits are used for selecting best hits for the gene  |
| <code>shortfac</code>     | a numeric that will scale up the chi-squared statistic before it is converted to short integer for storage in <code>ff</code> array  |
| <code>chrnames</code>     | character vector of chromosome identifiers, to be manipulated for certain query resolutions by the following parameters  |
| <code>smchrpref</code>    | prefix to convert <code>chrnames</code> into appropriate tokens for indexing <code>smlSet</code> elements as collected from the package named by parameter <code>smpack</code>   |
| <code>gchrpref</code>     | prefix to convert <code>chrnames</code> into appropriate tokens for obtaining gene metadata; in future this may need to be a string transformation function  |
| <code>schrpref</code>     | prefix to convert <code>chrnames</code> into appropriate tokens for use with <code>getSNPlocs</code> for the SNP location information package identified in <code>snpannopack</code> parameter below   |
| <code>geneApply</code>    | an <code>lapply</code> like function, defaults to <code>lapply</code>  |
| <code>geneannopk</code>   | character string, name of a <code>*.db</code> annotation package that annotates probe identifiers; or see <code>getCisMap</code> for additional possibilities concerning <code>FDb.*</code> complex token values for newer annotation formats                              |
| <code>snpannopk</code>    | character string, name of <code>SNPlocs.Hsapiens.dbSNP.*</code> package for obtaining; global function <code>snplocsDefault()</code> can be used to get a nominally current package name   |
| <code>smFilter</code>     | function accepting and returning an <code>smlSet-class</code> instance   |
| <code>SMFilterList</code> | list of functions, one element per <code>smlSet</code> package used in meta analysis, accepting and returning an <code>smlSet-class</code> instance  |
| <code>minchisq</code>     | threshold on test statistic value that must be met to include records on SNPs in the <code>All.cis.eQTLs</code> report   |
| <code>nperm</code>        | number of permutations to be used for plug-in FDR computation  |
| <code>useME</code>        | logical; if <code>TRUE</code> , use the rudimentary interface to the <code>MatrixEQTL</code> package from A. Shabalin on CRAN  |
| <code>maxfdr</code>       | Used in <code>All.cis.eQTLs</code> . The process of identifying “best” cis eQTL per probe leads to a probe-specific FDR. In <code>All.cis.eQTLs</code> we enumerate all probes and all SNP with FDR at most <code>maxfdr</code> , not just the best scoring SNP per probe. |
| <code>inbestcis</code>    | Used in <code>All.cis.eQTLs</code> . An instance of <code>mcwBestCis</code> that can be used to speed up the extraction of <code>All.cis.eQTL</code> .   |
| <code>smFilter4cis</code> | Used in <code>All.cis.eQTLs</code> . A function accepting and returning an <code>smlSet</code> instance. When <code>inbestcis</code> parameter is <code>NULL</code> , this filter will be used for identifying the best SNP per probe.                                     |



|               |   |
|---------------|---|
| smFilter4all  | Used in All.cis.eQTLs. A function accepting and returning an smlSet instance. This filter will be used for identifying the best SNP per probe. This filter should not affect the number of probes.  |
| x             | instance of mcwBestCis  |
| type          | character, either 'data.frame' or 'GRanges'   |
| excludeRadius | numeric, defaulting to NULL; if non-null, defines radius around gene region that is excluded for cis SNP scoring; must be less than radius  |
| keepMapCache  | logical, if TRUE, returned mcwBestCis object will include an environment loaded with chromosome-specific lists of maps from genes to cis SNP names; if FALSE, the mapCache environment returned will be empty – NB, this feature has been found to add too much volume to returned objects and is suspended...  |
| exFilter      | this function is passed to <a href="#">getSS</a> ; see Details  |
| exFilterList  | for metaanalytic applications, a list of functions in correspondence with the elements of smpackvec to be passed to <a href="#">getSS</a> ; see Details   |
| getDFFITS     | logical; a component storing max DFFITS value for each gene will be retained if this argument TRUE  |
| ...           | not used  |
| SSgen         | function to be used to create smlSet instance for testing – in general, GG-Base::getSS has been used to pull the ExpressionSet and SnpMatrix data from a named package, but in some cases a specialize task is needed to create the desired smlSet. Whatever is passed to SSgen must return an smlSet instance. |

## Details

geneApply can be set to `parallel::mclapply`, for example, in a multicore context.

mcwBestCis stands for 'multi-chromosome-wide best cis' eQTL report container.

It is possible that the filtering processes should be broken into genotype filtering and expression probe filtering.

`fdr(x)` will return a numeric vector of plug-in FDR estimates corresponding to probe:association tests as ordered in the fullreport of a \*Cis container. More metadata should be attached to the output of this function.

`exFilter` may seem redundant with `smFilter`, but its existence allows simpler management of multitissue expression archives (which may have several records per individual) with germ line genotype data (which will have only one record per individual). In this setting, use `exFilter` to select records for the tissue of interest; this will occur early in the smlSet generation process.

## Value

an instance of `mcwBestCis`

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
getClass("mcwBestCis")
## Not run:
best.cis.eQTLs(chrnames="20")

## End(Not run)
```

---

|                  |  |
|------------------|--|
| best.trans.eQTLs | <i>collect strongest trans SNP-gene associations in a buffer of size K genes per SNP</i> |
|------------------|--|

---

**Description**

collect strongest trans SNP-gene associations in a buffer of size K genes per SNP

**Usage**

```
best.trans.eQTLs(smpack, rhs, genechrnum, snpchrnum, K = 20,
  targdirpref = "tsco", batchsize = 200, radius = 2e+06, genequeryprefix = "",
  snploadprefix = "chr", snplocprefix = "chr", geneannopk, snpannopk,
  exFilter = function(x) x, smFilter = function(x) x,
  geneApply = lapply, SSgen = GGBase::getSS)
```

**Arguments**

|                 |  |
|-----------------|--|
| smpack          | character string naming a package from which <a href="#">smlSet-class</a> instances can be generated using <a href="#">getSS</a>   |
| rhs             | passed to <a href="#">snp.rhs.tests</a> for covariate or stratification adjustments; for permutation analysis, covariates should be handled via <a href="#">regressOut</a> |
| genechrnum      | character vector of chromosome identifiers for genes, typically as <code>character(1:22)</code> for somatic genes in human studies   |
| snpchrnum       | specific chromosome identifier for all SNP to be analyzed  |
| K               | the size of the buffer: scores will be recorded for the most strongly associated K genes for each SNP  |
| targdirpref     | character string where buffer data will be held in ff archives   |
| batchsize       | passed to <a href="#">ffrowapply</a> as scores are filtered from comprehensive testing to fill the buffer  |
| radius          | numeric: for same-chromosome tests, tests will not be performed for SNP-gene combinations with base-pair proximity smaller than radius                                     |
| genequeryprefix | string: used when the numeric chromosome identifier requires a prefix like 'chr' for annotation query resolution on gene location  |
| snploadprefix   | string: used when the package identified in smpack requires a prefix to the snpchrnum token for <a href="#">getSS</a> retrieval of <a href="#">smlSet</a> instance         |

|              |   |
|--------------|---|
| snplocprefix | string: used when the numeric chromosome identifier requires a prefix like 'chr' for annotation query resolution on SNP location  |
| geneannopk   | package to be used for CHRLOC and CHRLOCEND queries for genes   |
| snpannopk    | package to be used to resolve getSNPlocs calls  |
| exFilter     | function returning an smlSet instance, operating on expression component prior to smFilter application and eQTL testing   |
| smFilter     | function returning an smlSet instance, operating on the full smlSet   |
| geneApply    | lapply-like function, typically mclapply or the like  |
| SSgen        | function to be used to create smlSet instance for testing – in general, GG-Base::getSS has been used to pull the ExpressionSet and SnpMatrix data from a named package, but in some cases a specialize task is needed to create the desired smlSet. Whatever is passed to SSgen must return an smlSet instance. |

## Value

instance of [transManager-class](#)

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

## Examples

```
## Not run:
if (.Platform$OS.type != "windows") { # ff overwrites failing 5.IX.12
  nsFilter2 = function(sms, var.cutoff=.5) {
    alliq = apply(exprs(sms),1,IQR)
    qs = quantile(alliq,var.cutoff, na.rm=TRUE)
    sms[ which(alliq > qs), ]
  }
  thefilt = function(x) GTFfilter( nsFilter2( clipPCs(x, 1:10), var.cutoff=.95 ), lower=.05 )
  tfile = tempfile()
  tfold = dir.create(tfile)
  t1 = best.trans.eQTLs( "GGdata", ~1, as.character(20:22), "22",
    geneannopk="illuminaHumanv1.db", snpannopk= snplocsDefault(),
    smFilter=thefilt, snploadprefix="", snplocprefix="ch", targdirpref=tfile)
  tt1 = transTab(t1)
  tt1o = tt1[ order(tt1["sumchisq"], decreasing=TRUE), ][1:10,]
  tt1o
}

## End(Not run)
```

---

|         |   |
|---------|---|
| bindmaf | <i>bind testing metadata to a best.cis.eQTLs result</i> |
|---------|---|

---

## Description

bind testing metadata to a best.cis.eQTLs result

## Usage

```
bindmaf(smpack = "GGdata", smchr = "20", obj, SSgen=GGBase::getSS)
meta.bindmaf (smpackvec=c("GGdata", "hmyriB36"),
              smchr="20", obj, usemaxMAF=FALSE, SSgen=GGBase::getSS)
```

## Arguments

|           |  |
|-----------|--|
| smpack    | name of a package to which <a href="#">getSS</a> can be applied to generate an instance of <a href="#">smlSet-class</a>  |
| smpackvec | a vector of candidate smpack values for metaanalysis across populations or tissues   |
| smchr     | the chromosome name as used in the names of the <code>smlist</code> output for the <code>getSS</code> result   |
| obj       | an instance of <a href="#">mcwBestCis-class</a> generated using the package named in <code>smpack</code>   |
| usemaxMAF | if TRUE, label a SNP with maximum MAF observed across populations, otherwise compute the MAF for the combined genotypes across populations represented by the various <code>smlSet</code> instances generated with the <code>smpackvec</code> spec.  |
| SSgen     | function to be used to create <code>smlSet</code> instance for testing – in general, <code>GGBase::getSS</code> has been used to pull the <code>ExpressionSet</code> and <code>SnpMatrix</code> data from a named package, but in some cases a specialize task is needed to create the desired <code>smlSet</code> . Whatever is passed to <code>SSgen</code> must return an <code>smlSet</code> instance. |

## Details

computes the MAF of most highly associated SNP per gene, and distance between that SNP and the transcription limits of the gene, assigning 0 for this if the SNP lies within the transcription limits

## Value

a `GRanges` instance

## Note

This will be used to stratify the permuted scores.

**Examples**

```
## Not run:
b1 = best.cis.eQTLs(chr="20") # sharply filtered
b1b = bindmaf(obj=b1)

## End(Not run)
```

---

|                 |                          |
|-----------------|--------------------------|
| CisConfig-class | <i>Class "CisConfig"</i> |
|-----------------|--------------------------|

---

**Description**

Object specifying configuration of cis-eQTL search, to be used with All.cis

**Objects from the Class**

Objects can be created by calls of the form `new("CisConfig")`. Use replacement methods to update the fields.

**Slots**

**smpack:** character string identifying package holding the expression and genotype data; see [getSS](#)

**rhs:** Object of class "formula" right hand side for calls to [snp.rhs.tests](#)

**nperm:** Object of class "integer" number of permutations for plug in FDR

**folderStem:** Object of class "character" string used for scratch space folders, relative to current folder

**radius:** Object of class "integer" radius of search

**shortfac:** Object of class "integer" scores are scaled up by this factor so that precision can be retained in short integer representation

**chrnames:** Object of class "character" string identifying chromosome label used in gene annotation retrieval – typically length 1

**smchrpref:** Object of class "character" prefix to be attached to chromosome label in chrnames to pick out the element of [smlSet-class](#) instance used in testing

**gchrpref:** Object of class "character" prefix on chrnames token to be used for gene location retrievals

**schrpref:** Object of class "character" prefix on chrnames token to be used with [SNPlocs](#) package for retrieval of SNP locations

**geneApply:** Object of class "function" iterator over genes, could be `lapply` or `mclapply`

**geneannopk:** Object of class "character" Bioconductor annotation package for gene locations, typically for expression array

**snpannopk:** Object of class "character" Bioconductor dbSNP annotation package

**smFilter:** Object of class "function" function to be applied to `smlSet` instance that yields an `smlSet` instance with required contents; could apply MAF restriction for example by calling `MAFfilter`

**exFilter:** Object of class "function" function that is run right after smlSet is materialized, permitting replacement or filtering of expression data, when, for example, the ExpressionSet includes multiple tissue types

**keepMapCache:** Object of class "logical" for enhancing processing of gene-SNP cis mapping with a global cache

**SSgen:** Object of class "function" function that accepts name of an expression+SnpMatrix package (as generated by [externalize](#)), a chromosome tag (chrnames prefixed by smchrpref), and a function, and returns an smlSet instance

**excludeRadius:** Object of class "integerOrNULL" which will determine what interval about the gene is excluded for cis testing; 0 should exclude all within-gene SNP, but needs testing

**estimates:** Object of class "logical" if TRUE, estimates and standard errors (expanded and reduced in storage as a short int, using shortfac) are generated and retained

## Methods

**chrnames** signature(x = "CisConfig"): ...

**chrnames<-** signature(object = "CisConfig", value = "character"): ...

**estimates** signature(x = "CisConfig"): ...

**estimates<-** signature(object = "CisConfig", value = "logical"): ...

**excludeRadius** signature(x = "CisConfig"): ...

**excludeRadius<-** signature(object = "CisConfig", value = "integer"): ...

**exFilter** signature(x = "CisConfig"): ...

**exFilter<-** signature(object = "CisConfig", value = "function"): ...

**gchrpref** signature(x = "CisConfig"): ...

**gchrpref<-** signature(object = "CisConfig", value = "character"): ...

**geneannopk** signature(x = "CisConfig"): ...

**geneannopk<-** signature(object = "CisConfig", value = "character"): ...

**geneApply** signature(x = "CisConfig"): ...

**geneApply<-** signature(object = "CisConfig", value = "function"): ...

**initialize** signature(.Object = "CisConfig"): ...

**keepMapCache** signature(x = "CisConfig"): ...

**keepMapCache<-** signature(object = "CisConfig", value = "logical"): ...

**radius** signature(x = "CisConfig"): ...

**radius<-** signature(object = "CisConfig", value = "integer"): ...

**rhs** signature(x = "CisConfig"): ...

**rhs<-** signature(object = "CisConfig", value = "function"): ...

**schrpref** signature(x = "CisConfig"): ...

**schrpref<-** signature(object = "CisConfig", value = "character"): ...

**shortfac** signature(x = "CisConfig"): ...

**shortfac<-** signature(object = "CisConfig", value = "integer"): ...

```

show signature(object = "CisConfig"): ...
smchrpref signature(x = "CisConfig"): ...
smchrpref<- signature(object = "CisConfig", value = "character"): ...
smFilter signature(x = "CisConfig"): ...
smFilter<- signature(object = "CisConfig", value = "function"): ...
snpannopk signature(x = "CisConfig"): ...
snpannopk<- signature(object = "CisConfig", value = "character"): ...
SSgen signature(x = "CisConfig"): ...
SSgen<- signature(object = "CisConfig", value = "function"): ...

```

## Examples

```
showClass("CisConfig")
```

---

|              |                |
|--------------|----------------|
| cisRun-class | Class "cisRun" |
|--------------|----------------|

---

## Description

manage results of All.cis eQTL analysis

## Objects from the Class

Objects can be created by calls of the form `new("cisRun", ...)`.

## Slots

```

seqnames: Object of class "Rle" ~~
ranges: Object of class "IRanges" ~~
strand: Object of class "Rle" ~~
elementMetadata: Object of class "DataFrame" ~~
seqinfo: Object of class "Seqinfo" ~~
metadata: Object of class "list" ~~

```

## Extends

Class "[GRanges](#)", directly. Class "[GenomicRanges](#)", by class "[GRanges](#)", distance 2. Class "[Vector](#)", by class "[GRanges](#)", distance 3. Class "[GenomicRangesORmissing](#)", by class "[GRanges](#)", distance 3. Class "[GenomicRangesORGRangesList](#)", by class "[GRanges](#)", distance 3.

Class "[Annotated](#)", by class "[GRanges](#)", distance 4.

## Methods

No methods defined with class "cisRun" in the signature.

**Note**

intent is to simplify output of cis eQTL testing in a GRanges instance

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
showClass("cisRun")
```

---

|             |   |
|-------------|---|
| collectBest | <i>given a collection of All.cis outputs (cisRun instances) compute FDRs for various filterings</i> |
|-------------|---|

---

**Description**

given a collection of All.cis outputs (cisRun instances) compute FDRs for various filterings

**Usage**

```
collectBest(fns,
  targetname = "harvest",
  mafs = c(0.01, 0.02, 0.025, 0.03333, 0.05, 0.075, 0.1),
  hidists = c(10000, 25000, 50000, 75000, 1e+05, 250000), interimSaves=FALSE)

collectFiltered( fns, targetname="harvest",
  mafs = c(.01, .02, .025, .03333, .05, .075, .1),
  hidists = c(10000, 25000, 50000, 75000, 100000, 250000),
  filterFun = cis.FDR.filter.best, filtApplier=lapply,
  interimSaves=FALSE)
```

**Arguments**

|              |  |
|--------------|--|
| fns          | names of .rda with the cisRun outputs                  |
| targetname   | basename of rda file to be emitted                     |
| mafs         | lower bounds on MAF for filtering                      |
| hidists      | upper bounds on cis radius for filtering               |
| filterFun    | function like GGtools:::cis.FDR.filter.best            |
| filtApplier  | function like lapply                                   |
| interimSaves | logical, if TRUE save list at each maf/dist transition |

**Details**

[pifdr](#) is repeatedly used to generate conditional plugin FDR for different filtering criteria



**Value**

a list of lists is written to disk incrementally, as the job can be long running

**Note**

This is the workhorse of sensitivity analysis. Permits counting of genes with eQTL at selected FDR for various criteria on cis radius and MAF.

**Examples**

```
## Not run:
#
# contents of fns are two chromosomes of cis runs for CEU
#
fns = dir(system.file("rdas", package="GGtools"), full=TRUE)
cc = collectBest(fns, maf=c(.01, .05), hidists=c(10000, 50000))
sapply(cc, sapply, function(x) sum(x$fdr <= 0.01))
#
# this tells us which to keep
#
kp = cc[["0.05"]][["50000"]]
kp = kp[kp$fdr <= 0.01,]
#
# the hits are in the table above; the following function
# retrieves the initial scores giving rise to the filtered
# hits
#
pullHits = function(fns, atts) {
  tmp = lapply(fns, function(x) get(load(x)))
  k1 = lapply(tmp, function(x) paste(names(x), x$snp, sep=":"))
  atk = paste(atts$genes, atts$bestsnp, sep=":")
  tmp = lapply(1:length(tmp), function(x) tmp[[x]][ match( atk, k1[[x]], nomatch=0 ) ])
  curans = do.call(c, lapply(tmp, as, "GRanges"))
  newword = match( atk, paste(names(curans), curans$snp, sep=":"))
  newfdr = atts$fdr[newword]
  curans$fdr = newfdr
  curans
}
pullHits( fns, kp )
#
#
#
# after executing code in example for All.cis (protected by dontrun)
# and running save(f1, file="f1.rda"), the following will work
# genewise max score
cf1 = collectFiltered("f1.rda", maf=.02, hidists=25000, targetname="gwise")
# SNPwise scores, all
cf2 = collectFiltered("f1.rda", maf=.02, hidists=25000, targetname="swise",
  filterFun = cis.FDR.filter.SNPcentric.complete )
# SNPwise scores, best per SNP when SNP is cis to multiple genes
cf3 = collectFiltered("f1.rda", maf=.02, hidists=25000, targetname="swise2",
  filterFun = cis.FDR.filter.SNPcentric )
```

```
## End(Not run) # end dontrun
```

---

|           |  |
|-----------|--|
| eqtlTests | <i>compute association statistics between all probes and SNP in an smlSet instance</i> |
|-----------|--|

---

### Description

compute association statistics (or point estimates and standard errors) between all probes and SNP in an smlSet instance, using out-of-memory storage

### Usage

```
eqtlTests(smlSet, rhs = ~1 - 1, runname = "foo",
  targdir = "foo", geneApply = lapply,
  shortfac = 100,
  checkValid = TRUE, useUncertain = TRUE,
  glmfamily = "gaussian")
```

```
eqtlEstimates(smlSet, rhs = ~1 - 1, runname = "foo",
  targdir = "foo", geneApply = lapply,
  shortfac = 10000,
  checkValid = TRUE, useUncertain = TRUE,
  glmfamily = "gaussian")
```

### Arguments

|              |   |
|--------------|---|
| smlSet       | instance of <a href="#">smlSet</a>  |
| rhs          | fragment of a standard formula, minus a dependent variable (i.e., starts with tilde); bindings will be sought in <code>pData(smlSet)</code>                           |
| runname      | string used to identify output ff files   |
| targdir      | string naming the folder where ff outputs will reside   |
| geneApply    | analog to <code>lapply</code> to drive iteration over probes  |
| shortfac     | ff contents will be multiplied by this quantity and stored as short integers  |
| checkValid   | logical, will apply <code>validObject</code> to <code>smlSet</code> if TRUE   |
| useUncertain | logical, passed as <code>uncertain</code> parameter to <a href="#">snp.rhs.tests</a> to specify whether uncertain genotypes will be used (as 'dosage' in GLM fitting) |
| glmfamily    | family specification for <a href="#">snp.rhs.tests</a>  |

**Details**

The purpose of the eqtlTests function is to allow very substantial eQTL search processes to occur with R. For several million SNP and tens of thousands of probes, the storage of test results requires attention to parsimony. The storage occurs out of memory, using the ff package, and employs short integers to represent chi squared statistics. These are scaled up prior to storage, and will be scaled down prior to use.

eqtlEstimates will use compact storage for both the point estimates and standard errors of association estimated under an additive genetic model

**Value**

returns an instance of eqtlTestsManager

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
hm2ceuSMS = getSS("GGtools", c("20"), renameChrs=c("chr20"))
library(illuminaHumanv1.db)
cptag = get("CPNE1", revmap(illuminaHumanv1SYMBOL))
indc = which(featureNames(hm2ceuSMS) == cptag[1])
#
# get a set of additional genes on chr20
all20 = get("20", revmap(illuminaHumanv1CHR))
g20 = unique(c(all20[1:10], cptag))
#
hm = hm2ceuSMS[probeId(g20),] # reduce problem
td = tempdir()
curd = getwd()
setwd(td)
time.lapply = unix.time(e1 <- eqtlTests( hm, ~male ))
time.lapply
e1
# best chisq(1) for CPNE1
topFeats(probeId(cptag), e1)
setwd(curd)
```

---

eqtlTestsManager-class

*Class "eqtlTestsManager"*

---

**Description**

manage out-of-memory elements of an eQTL search

### Objects from the Class

Objects can be created by calls of the form `new("eqtlTestsManager", ...)`.

### Slots

**fffile:** Object of class "ff\_matrix" chisquared statistics stored as short ints in ff out of memory file

**call:** Object of class "call" audit of creation call

**sess:** Object of class "ANY" session info structure at time of creation

**exdate:** Object of class "ANY" date at time of creation

**shortfac:** Object of class "numeric" number by which chisq stats are multiplied to allow recovery of precision

**geneanno:** Object of class "character" string naming annotation package relevant for probe identifier translation

**df:** Object of class "numeric" degrees of freedom of chisq stats

**summaryList:** Object of class "list" list of genotype statistical summaries

### Methods

[ signature(x = "eqtlTestsManager", i = "ANY", j = "ANY", drop = "ANY"): extract chisq statistics properly rescaled from short int to double

**show** signature(object = "eqtlTestsManager"): concise report

**topFeats** signature(feats = "probeId", mgr = "eqtlTestsManager"): extract highest scores for SNP associated with given probeId

**topFeats** signature(feats = "rsid", mgr = "eqtlTestsManager"): extract highest scores for probes associated with given SNP

### Note

instances are created by [eqtlTests](#)

### Author(s)

VJ Carey <stvjc@channing.harvard.edu>

### Examples

```
showClass("eqtlTestsManager")
```

ex

*ExpressionSet instance for illustrating integrative smlSet container***Description**

ExpressionSet instance for illustrating integrative smlSet container

**Usage**

data(eset)

**Format**

The format is: Formal class 'ExpressionSet' [package "Biobase"] with 7 slots ..@ experimentData :Formal class 'MIAME' [package "Biobase"] with 13 slots

```

.. ..@ name : chr ""
.. ..@ lab : chr ""
.. ..@ contact : chr ""
.. ..@ title : chr ""
.. ..@ abstract : chr ""
.. ..@ url : chr ""
.. ..@ pubMedIds : chr ""
.. ..@ samples : list()
.. ..@ hybridizations : list()
.. ..@ normControls : list()
.. ..@ preprocessing : list()
.. ..@ other : list()
.. ..@ .__classVersion__:Formal class 'Versions' [package "Biobase"] with 1 slots
.. .. .@ .Data:List of 2
.. .. . .$. : int [1:3] 1 0 0
.. .. . .$. : int [1:3] 1 1 0
.. ..@ assayData :<environment: 0x10bf12948>
.. ..@ phenoData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots
.. .. .@ varMetadata :'data.frame': 7 obs. of 1 variable:
.. .. . .$. labelDescription: chr [1:7] "hapmap family id" "hapmap person id" "id of mother of this
person" "id of father of this person" ...
.. .. .@ data :'data.frame': 90 obs. of 7 variables:
.. .. . .$. famid : int [1:90] 1341 1341 1341 1340 1340 1340 1340 1341 1341 ...
.. .. . .$. persid : int [1:90] 14 2 13 9 10 2 11 1 11 1 ...
.. .. . .$. mothid : int [1:90] 0 14 0 0 0 12 0 10 0 12 ...
.. .. . .$. fathid : int [1:90] 0 13 0 0 0 11 0 9 0 11 ...
.. .. . .$. sampid : Factor w/ 90 levels "NA06985","NA06991",...: 1 2 3 4 5 6 7 8 9 10 ...
.. .. . .$. isFounder: logi [1:90] TRUE FALSE TRUE TRUE TRUE FALSE ...
.. .. . .$. male : logi [1:90] FALSE FALSE TRUE TRUE FALSE FALSE ...
.. .. .@ dimLabels : chr [1:2] "sampleNames" "sampleColumns"
.. .. .@ .__classVersion__:Formal class 'Versions' [package "Biobase"] with 1 slots
.. .. . .@ .Data:List of 1

```

```

.. .. .. .. .$ : int [1:3] 1 1 0
..@ featureData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots
.. .. ..@ varMetadata :'data.frame': 0 obs. of 1 variable:
.. .. .. .$ labelDescription: chr(0)
.. .. ..@ data :'data.frame': 47293 obs. of 0 variables
.. .. ..@ dimLabels : chr [1:2] "featureNames" "featureColumns"
.. .. ..@ __classVersion__:Formal class 'Versions' [package "Biobase"] with 1 slots
.. .. .. ..@ .Data:List of 1
.. .. .. .. .$ : int [1:3] 1 1 0
..@ annotation : chr "illuminaHumanv1.db"
..@ protocolData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots
.. .. ..@ varMetadata :'data.frame': 0 obs. of 1 variable:
.. .. .. .$ labelDescription: chr(0)
.. .. ..@ data :'data.frame': 90 obs. of 0 variables
.. .. ..@ dimLabels : chr [1:2] "sampleNames" "sampleColumns"
.. .. ..@ __classVersion__:Formal class 'Versions' [package "Biobase"] with 1 slots
.. .. .. ..@ .Data:List of 1
.. .. .. .. .$ : int [1:3] 1 1 0
..@ __classVersion__:Formal class 'Versions' [package "Biobase"] with 1 slots
.. .. ..@ .Data:List of 4
.. .. .. .$ : int [1:3] 2 14 0
.. .. .. .$ : int [1:3] 2 13 7
.. .. .. .$ : int [1:3] 1 3 0
.. .. .. .$ : int [1:3] 1 0 0

```

## Details

Expression data harvested in 2007 from GENEVAR

[ftp://ftp.sanger.ac.uk/pub/genevar/CEU\\_parents\\_norm\\_march2007.zip](ftp://ftp.sanger.ac.uk/pub/genevar/CEU_parents_norm_march2007.zip)

## Examples

```
data(eset) # yields ExpressionSet instance called ex
```

---

getCisMap

*create, using Bioconductor annotation resources, a structure that enumerates SNP in the vicinity of ('cis' to) genes*

---

## Description

create a structure that enumerates SNP in the vicinity of ('cis' to) genes

**Usage**

```
getCisMap(radius = 50000, gchr = "20",
          schr = "ch20", geneannopk = "illuminaHumanv1.db",
          snpannopk = snplocsDefault(),
          as.GRangesList = FALSE, excludeRadius=NULL)
```

**Arguments**

|                |  |
|----------------|--|
| radius         | How far, in bases, up or down stream from the asserted coding region limits to include SNP   |
| gchr           | the token to be used to acquire locations for probes on a specified chromosome, using revmap([dbpk]CHR)  |
| schr           | the token to be used to acquire locations for SNP on a specified chromosome, using getSnplocs  |
| geneannopk     | character string naming a Bioconductor .db expression chip annotation package; or a complex string with first part naming a Bioconductor FDb.* annotation package, colon separator, and a second string naming the getter hook that when called returns a GRanges with names corresponding to features and ranges corresponding to feature extents. For example "FDb.InfiniumMethylation.hg19:get27k" is valid. Note that in this case, gchr must have prefix "chr". |
| snpannopk      | character string naming a Bioconductor SNPlocs.* SNP metadata package  |
| as.GRangesList | logical telling whether a GRangesList should be returned   |
| excludeRadius  | numeric or NULL: radius of interval around gene extent from which SNP will be excluded, required to be less than radius  |

**Details**

This is a utility that the developer would rather not export. The complexity of harmonizing queries among probe and SNP annotation resources leads to a somewhat fragile product. Users who have their own gene ranges and SNP locations can examine the namelist component of the output of the default call to see what is expected for the \*.cis.eQTLs function. For the set of chromosomes to be analyzed, there will be a list of chromosome specific namelist-like lists.

**Value**

Instance of cisMap class, which will retain SNP location, gene range, and radius information for auditing.

**Examples**

```
## Not run:
getCisMap()

## End(Not run)
```

---

|            |  |
|------------|--|
| gwSnpTests | <i>execute a series of tests for association between genotype and expression</i> |
|------------|--|

---

### Description

execute a series of tests for association between genotype and expression

### Usage

```
gwSnpTests(sym, sms, ...)  
topSnps(x, n=10)
```

### Arguments

|     |   |
|-----|---|
| sym | instance of <a href="#">probeId</a> or <a href="#">genesym</a>                              |
| sms | instance of <a href="#">smlSet-class</a>  |
| x   | instance of <a href="#">gwSnpScreenResult</a>   |
| n   | integer, number of test results to be reported, sorted decreasing from the most significant |
| ... | not used  |

### Details

The plot method for [gwSnpScreenResult](#) instances takes a second argument, the name of a Bioconductor `SNPlocs.*` package.

### Value

an instance of the [gwSnpScreenResult](#) class, to be examined by `topSnps`

### Note

The most basic application yields one d.f. chi-squared statistics based on score tests.

### Author(s)

VJ Carey <stvjc@channing.harvard.edu>

### Examples

```
s20 = getSS("GGtools", "20")  
t1 = gwSnpTests(genesym("CPNE1")~male, s20)  
topSnps(t1)  
## Not run:  
plot(t1, snplocsDefault())  
  
## End(Not run)
```



---

pifdr

*utility for computing plug-in FDR*

---

### Description

utility for computing plug-in FDR

### Usage

```
pifdr( obs, perms, npts=1999, applier=sapply)
```

### Arguments

|         |  |
|---------|--|
| obs     | observed association scores  |
| perms   | vector of association scores under permutation; length should be integer multiple of length(obs)                         |
| npts    | number of points spanning the range of obs to be used for a lossy grid-based computation; only used if length(obs)>npts. |
| applier | function that iterates the computation   |

### Details

As currently implemented the algorithm is quadratic in length(obs). While it is possible to get a unique FDR value for every element of obs, an approximate approach yields practically identical precision and by default this will be used for obs with length 2000 or more. In this case, [approx](#) is used with rule=2 to interpolate from the grid-based FDR estimates back to the data values.

### Value

vector of plug-in FDR estimates congruent to obs

### References

Hastie Tibshirani and Friedman Elements of Statistical Learning ch 18.7

### Examples

```
X = rchisq(100,1)
Y = rchisq(300,1)
qqplot(pifdr(X,Y), rchisq(100,1))
```

---

|          |  |
|----------|--|
| richNull | <i>bind metadata concerning SNP allele frequency and other aspects of optimized cis-eQTL association to an mcwBestCis instance</i> |
|----------|--|

---

### Description

bind metadata concerning SNP allele frequency and other aspects of optimized cis-eQTL association to an mcwBestCis instance, to allow conditional FDR computation

### Usage

```
richNull(..., MAF1b = 0.01, npc = 10, radius = 250000, nperm = 1,
  innerFilt = function(x) x, outerFilt = function(x) x)

meta.richNull(..., MAF1b=.01, npc=10, radius=250000,
  nperm=1, innerFilt=function(x)x, outerFilt=function(x)x)
#
# internally:
#
# bigfilt = function(z)
#   outerFilt(MAFfilter(clipPCs(permEx(innerFilt(z))), 1:npc), lower=MAF1b))
#
```

### Arguments

|           |   |
|-----------|---|
| ...       | should provide bindings for smpack and chrnames, which will be used to obtain gene/probe locations; see <a href="#">getSS</a> for information on smpack settings. meta.richNull allows a vector of smpack values bound to smpackvec |
| MAF1b     | lower bound on SNP MAF for null distribution evaluation   |
| npc       | number of expression principal components to be removed   |
| radius    | radius used for testing   |
| nperm     | This establishes how many permutations of expression against genotype will be performed for this process.   |
| innerFilt | function immediately applied to generated smlSet instances  |
| outerFilt | function applied to generated smlSet instances after clipPCs and MAFfilter are applied in that order  |

### Details

The purpose of richNull is to obtain realizations from the permutation distribution of cis-eQTL association statistics, binding information on the characteristics of the optimal results with the scores. This allows us to use conditioning with the realizations from the permutation distribution.

### Value

richNull returns a list of nperm mcwBestCis instances with additional metadata bound in

**Author(s)**

Vince Carey <stvjc@channing.harvard.edu>

---

|          |  |
|----------|--|
| sensanal | <i>Summarize information from a collection of eQTL searches for sensitivity assessment</i> |
|----------|--|

---

**Description**

Summarize information from a collection of eQTL searches for sensitivity assessment

**Usage**

```
sensanal(object, fdrbound)
```

**Arguments**

|          |   |
|----------|---|
| object   | instance of <a href="#">sensiCisInput-class</a>           |
| fdrbound | numeric upper bound on FDR for declarations of eQTL yield |

**Details**

Sensitivity analysis for cis-eQTL search involves checking effects of scope of search, allele frequency filtering, and adjustment for expression heterogeneity on eQTL declarations. In this version, we focus on collections of outputs of [best.cis.eQTLs](#), to which the values of tuning parameters are bound. These collections are identified in a [sensiCisInput-class](#) instance, and the `sensanal` function processes these outputs into a [sensiCisOutput-class](#) instance for tabulation and visualization.

**Value**

a [sensiCisOutput-class](#) instance

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

---

sensiCisInput-class    *Class "sensiCisInput"*

---

### Description

Manage references to collections of cis-eQTL searches for sensitivity analysis.

### Objects from the Class

Objects can be created by calls of the form `new("sensiCisInput", ...)`.

### Slots

**cisMgrFiles:** Object of class "character": a vector of filenames, each file is an instance of class [mcwBestCis-class](#)

**cisMgrProperties:** Object of class "list" one vector with named elements per element of `cisMgrFiles`, with components `rad`, `excl`, `maf`, `nperm`, `npc`; see details below.

**probeannopk:** Object of class "character", identifying a bioconductor probe annotation package that can be used to map probe identifiers to other vocabularies or feature value sets

### Methods

**sensanal** signature(object = "sensiCisInput", fdrbound = "numeric"): generates an instance of [sensiCisOutput-class](#) with summarization of sensitivities

**show** signature(object = "sensiCisInput"): concise rendering

### Note

This version of sensitivity analysis support is rudimentary and involves manual construction of metadata that should be extractable from analysis outputs. The radius of the cis search (and radius of excluded interior if used) are identified as elements named `rad` and `excl` in the `cisMgrProperties` vectors; additional elements `maf`, `nperm`, and `npc` define the lower bound for minor allele frequency, number of permutations for plug-in FDR computation, and number of principal components removed to adjust for expression heterogeneity in the associated cis-eQTL search.

### Examples

```
showClass("sensiCisInput")
```

---

sensiCisOutput-class    *Class* "sensiCisOutput"

---

### Description

This class helps to manage the results from a collection of cis-eQTL searches.

### Objects from the Class

Objects can be created by calls of the form `new("sensiCisOutput", ...)`.

### Slots

**byGene:** Object of class "GRanges", organized to provide ranges for genes and their best associated cis SNP

**bySNP:** Object of class "GRanges" organized to provide easy access to genomic coordinates of SNP found to be most strongly associated with a gene in cis

**tabAtFDRB:** Object of class "ANY" a flattened table that defines tuning parameters and eQTL yield for a collection of searches

**input:** Object of class "sensiCisInput" : object that describes the files and parameter settings used for the sensitivity analysis

**thecall:** Object of class "call": the call generating this instance

**fdrbound:** Object of class "numeric": gives the upper bound on FDR for declaring an eQTL

**sessionInfo:** Object of class "ANY": describes state of system in which the object was made.

### Methods

**show** signature(object = "sensiCisOutput"): concise rendering with hints

### Author(s)

VJ Carey <stvjc@channing.harvard.edu>

### Examples

```
showClass("sensiCisOutput")
```

---

|                |  |
|----------------|--|
| snplocsDefault | <i>name the default SNPlocs.Hsapiens.dbSNP.* package</i> |
|----------------|--|

---

**Description**

generate a string naming the default SNPlocs.Hsapiens.dbSNP.\* package for use with GGtools

**Usage**

```
snplocsDefault()
```

**Details**

allows centralized specification of SNPlocs resource package

**Value**

a character string, see example

**Examples**

```
snplocsDefault()
```

---

|             |   |
|-------------|---|
| strMultiPop | <i>serialization of a table from Stranger's multipopulation eQTL report</i> |
|-------------|---|

---

**Description**

serialization of a table from Stranger's multipopulation eQTL report

**Usage**

```
data(strMultiPop)
```

**Format**

A data frame with 39649 observations on the following 12 variables.

rsid a factor with levels rs...

genesym a factor with levels 37865 39692 ABC1 ABCD2 ABHD4 ACAS2 ...

illv1pid a factor with levels GI\_10047105-S GI\_10092611-A GI\_10190705-S GI\_10567821-S  
GI\_10835118-S GI\_10835186-S ...

snpChr a numeric vector

snpCoordB35 a numeric vector

probeMidCoorB35 a numeric vector

snp2probe a numeric vector  
 minuslog10p a numeric vector  
 adjR2 a numeric vector  
 assocGrad a numeric vector  
 permThresh a numeric vector  
 popSet a factor with levels CEU-CHB-JPT CEU-CHB-JPT-YRI CHB-JPT

### Details

imported from the PDF(!) distributed by Stranger et al as supplement to PMID 17873874

### Source

PMID 17873874 supplement

### References

PMID 17873874 supplement

### Examples

```
data(strMultiPop)
strMultiPop[1:2,]
```

---

transManager-class      *Class* "transManager"

---

### Description

simple container for manager of transScores output

### Objects from the Class

Objects can be created by calls of the form `new("transManager", ...)`.

### Slots

**base:** Object of class "list" includes ff references for scores and indices of genes corresponding to scores, and other metadata about the run

### Methods

**show** signature(object = "transManager"): simple reporter

### See Also

[transTab](#)

**Examples**

```
showClass("transManager")
```

---

|             |  |
|-------------|--|
| transScores | <i>obtain the top trans associations for each SNP in an smlSet</i> |
|-------------|--|

---

**Description**

obtain the top trans associations for each SNP in an smlSet

**Usage**

```
transScores(smpack, snpchr = "chr1", rhs, K = 20, targdirpref = "tsco", geneApply = lapply,
  chrnames = paste("chr", as.character(1:22), sep = ""), geneRanges = NULL, snpRanges = NULL,
  radius = 2e+06, renameChrs = NULL, probesToKeep = NULL, batchsize = 200,
  genegran = 50, shortfac = 10, wrapperEndo = NULL,
  geneannopk = "illuminaHumanv1.db",
  snpannopk = snplocsDefault(), gchrpref = "",
  schrpref = "ch", exFilter=function(x)x,
  SSgen=GGBase::getSS)
```

```
meta.transScores (smpackvec = c("GGdata", "hmyriB36"),
  snpchr = "22", rhsList=list(~1, ~1), K = 20, targdirpref = "mtsco",
  geneApply = lapply, chrnames = as.character(21:22),
  radius = 2e+06, renameChrs=NULL,
  probesToKeep=NULL, batchsize=200, genegran=50, shortfac=10, wrapperEndo=NULL,
  geneannopk = "illuminaHumanv1.db", snpannopk = snplocsDefault(),
  gchrpref = "", schrpref="ch",
  exFilterList= list(function(x)x, function(x)x),
  SMFilterList = list(function(x)x, function(x)x),
  SSgen = GGBase::getSS)
```

**Arguments**

|           |   |
|-----------|---|
| smpack    | name of package holding eset.rda providing 'ex' ExpressionSet when loaded, and holding SnpMatrix instances in inst/parts                                  |
| smpackvec | vector of names of package holding eset.rda providing 'ex' ExpressionSet when loaded, and holding SnpMatrix instances in inst/parts                       |
| snpchr    | name or vector of chromosome names of SNPs of interest  |
| rhs       | right hand side of snp.rhs.tests model for which expression is left hand side, e.g., covariates other than genotype                                       |
| rhsList   | list of right hand side of snp.rhs.tests model for which expression is left hand side, e.g., covariates other than genotype, one per element of smpackvec |
| K         | number of most highly associated features to be retained  |



|              |  |
|--------------|--|
| targdirpref  | prefix of target folder name (passed to <a href="#">eqtlTests</a>  |
| geneApply    | passed to <a href="#">eqtlTests</a>  |
| chrnames     | names of chromosomes harboring genes that will be tested for association with genotype   |
| geneRanges   | list of <a href="#">GRanges-class</a> instances containing chromosomal coordinate defined regions occupied by genes, with regions partitioned by chromosomes, and list element names as given in chrnames above  |
| snpRanges    | list of <a href="#">GRanges-class</a> instances with SNP addresses   |
| radius       | radius within which an association is considered cis and therefore the corresponding test statistic is set to zero   |
| renameChrs   | passed to <a href="#">getSS</a>  |
| probesToKeep | passed to <a href="#">getSS</a>  |
| batchsize    | defines batch size for <a href="#">ffrowapply</a>  |
| genegran     | passed to <a href="#">eqtlTests</a>  |
| shortfac     | passed to <a href="#">eqtlTests</a>  |
| wrapperEndo  | a function accepting and returning an <a href="#">smlSet</a> instance, evaluated before numerical analysis of associations, which will be executed on the output of this function  |
| gchrpref     | prefix to convert chrnames into appropriate tokens for obtaining gene metadata; in future this may need to be a string transformation function   |
| schrpref     | prefix to convert chrnames into appropriate tokens for use with <a href="#">getSNPlocs</a> for the SNP location information package identified in <a href="#">snpannopack</a> parameter below  |
| geneannopk   | character string naming a Bioconductor .db expression chip annotation package  |
| snpannopk    | character string naming a Bioconductor <a href="#">SNPlocs.*</a> SNP metadata package  |
| exFilter     | function to transform <a href="#">ExpressionSet</a> component of retrieved <a href="#">smlSet</a> , to reduce probe sets in use, for example   |
| exFilterList | list of functions serving as <a href="#">exFilters</a> for each of the elements of <a href="#">smpackvec</a>   |
| SMFilterList | list of functions servicing as <a href="#">wrapperEndos</a> for each of the elements of <a href="#">smpackvec</a>  |
| SSgen        | function to be used to create <a href="#">smlSet</a> instance for testing – in general, <a href="#">GG-Base::getSS</a> has been used to pull the <a href="#">ExpressionSet</a> and <a href="#">SnpMatrix</a> data from a named package, but in some cases a specialize task is needed to create the desired <a href="#">smlSet</a> . Whatever is passed to <a href="#">SSgen</a> must return an <a href="#">smlSet</a> instance. |

**Value**

a list with elements

|        |   |
|--------|---|
| scores | an S by K ff matrix where S is number of SNPs, K is number of best features to be retained, with element s,k the kth largest score statistic among association tests computed for SNP s |
| inds   | an S by K ff matrix with s,k element telling which element of <a href="#">guniv</a> (see below) is the gene giving the kth largest score statistic for association                      |

guniv            the vector of gene identifiers defining the universe of genes tested  
 snpnames        vector of SNP identifiers  
 call             the call used to create the result

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
## Not run:
library(GGdata)
# need to define the geneRanges and snpRanges ...
transScores("GGdata", "20", renameChrs="chr20", chrnames="chr21")

## End(Not run)
```

---

|          |  |
|----------|--|
| transTab | <i>tabulate results of transScores run</i> |
|----------|--|

---

**Description**

tabulate results of transScores run

**Usage**

```
transTab(x, snps2keep, ...)
```

**Arguments**

x                a transManager instance.  
 snps2keep        character vector used for filtering snps whose scores will be retained; intersection with snps named in x will be used.  
 ...              not used

**Value**

data.frame instance

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

---

|        |   |
|--------|---|
| vcf2sm | <i>generate a SnpMatrix instance on the basis of a VCF (4.0) file</i> |
|--------|---|

---

### Description

generate a SnpMatrix instance on the basis of a VCF (4.0) file.

### Usage

```
vcf2sm(tbxfi, ..., gr, nmetacol)
```

### Arguments

|          |  |
|----------|--|
| tbxfi    | instance of <a href="#">TabixFile-class</a>                                  |
| ...      | not used   |
| gr       | instance of <a href="#">GRanges-class</a>                                    |
| nmetacol | numeric: tells number of columns used in each record as locus-level metadata |

### Details

This function is relevant only for diallelic SNP. If any base call is denoted '.', the associated genotype is set to missing (raw 0), even if the nonmissing call is ALT, implying at least one ALT.

### Value

an instance of [SnpMatrix-class](#)

### Author(s)

VJ Carey <stvjc@channing.harvard.edu>

### References

[http://www.1000genomes.org/wiki/doku.php?id=1000\\_genomes:analysis:vcf4.0](http://www.1000genomes.org/wiki/doku.php?id=1000_genomes:analysis:vcf4.0)

### Examples

```
# SRC: ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/pilot_data/release/2010_07/exon/CEU.exon.2010_03.genotypes.vcf.  
vref = system.file("vcf/CEU.exon.2010_09.genotypes.vcf.gz", package="GGtools")  
gg = GenomicRanges::GRanges(seqnames="1", IRanges::IRanges(10e6,20e6))  
vcf2sm(Rsamtools::TabixFile(vref), gr=gg, nmetacol=9L)
```

# Index

## \*Topic **classes**

- CisConfig-class, 13
- cisRun-class, 15
- eqtlTestsManager-class, 19
- sensiCisInput-class, 28
- sensiCisOutput-class, 29
- transManager-class, 31

## \*Topic **datasets**

- b1, 4
- ex, 21
- strMultPop, 30

## \*Topic **models**

- All.cis, 3
- best.cis.eQTLs, 6
- best.trans.eQTLs, 10
- bindmaf, 12
- collectBest, 16
- eqtlTests, 18
- getCisMap, 22
- gwSnpTests, 24
- pifdr, 25
- richNull, 26
- sensanal, 27
- snplocsDefault, 30
- transScores, 32
- transTab, 34
- vcf2sm, 35

## \*Topic **package**

- GGtools-package, 2

- [,eqtlTestsManager, ANY, ANY, ANY-method  
(eqtlTestsManager-class), 19

- All.cis, 3

- All.cis.eQTLs (best.cis.eQTLs), 6

- allSigCis-class (best.cis.eQTLs), 6

- Annotated, 15

- approx, 25

- b1, 4

- b2 (b1), 4

- best.cis.eQTLs, 6, 27

- best.trans.eQTLs, 10

- bindmaf, 12

- chrFilter (All.cis), 3

- chrnames (CisConfig-class), 13

- chrnames, CisConfig-method  
(CisConfig-class), 13

- chrnames<- (CisConfig-class), 13

- chrnames<-, CisConfig, character-method  
(CisConfig-class), 13

- chromsUsed (best.cis.eQTLs), 6

- chromsUsed, mcwBestCis-method  
(best.cis.eQTLs), 6

- cis.FDR.filter.best (collectBest), 16

- cis.FDR.filter.SNPcentric  
(collectBest), 16

- CisConfig-class, 13

- cisRun-class, 15

- collectBest, 16

- collectFiltered (collectBest), 16

- eqtlEstimates (eqtlTests), 18

- eqtlEstimatesManager-class  
(eqtlTestsManager-class), 19

- eqtlTests, 18, 20, 33

- eqtlTestsManager-class, 19

- estimates (CisConfig-class), 13

- estimates, CisConfig-method  
(CisConfig-class), 13

- estimates<- (CisConfig-class), 13

- estimates<-, CisConfig, logical-method  
(CisConfig-class), 13

- ex, 21

- excludeRadius (CisConfig-class), 13

- excludeRadius, CisConfig-method  
(CisConfig-class), 13

- excludeRadius<- (CisConfig-class), 13

- excludeRadius<-, CisConfig, integer-method  
(CisConfig-class), 13

- excludeRadius<- ,CisConfig, integerOrNULL-method  
(CisConfig-class), 13
- exFilter (CisConfig-class), 13
- exFilter, CisConfig-method  
(CisConfig-class), 13
- exFilter<- (CisConfig-class), 13
- exFilter<- ,CisConfig, function-method  
(CisConfig-class), 13
- externalize, 14
  
- fdr (best.cis.eQTLs), 6
- ffrowapply, 10, 33
- folderStem (CisConfig-class), 13
- folderStem, CisConfig-method  
(CisConfig-class), 13
- folderStem<- (CisConfig-class), 13
- folderStem<- ,CisConfig, character-method  
(CisConfig-class), 13
- fullreport (best.cis.eQTLs), 6
- fullreport, mcwBestCis, character-method  
(best.cis.eQTLs), 6
- fullreport, mcwBestCis, missing-method  
(best.cis.eQTLs), 6
  
- gchrpref (CisConfig-class), 13
- gchrpref, CisConfig-method  
(CisConfig-class), 13
- gchrpref<- (CisConfig-class), 13
- gchrpref<- ,CisConfig, character-method  
(CisConfig-class), 13
- geneannopk (CisConfig-class), 13
- geneannopk, CisConfig-method  
(CisConfig-class), 13
- geneannopk<- (CisConfig-class), 13
- geneannopk<- ,CisConfig, character-method  
(CisConfig-class), 13
- geneApply (CisConfig-class), 13
- geneApply, CisConfig-method  
(CisConfig-class), 13
- geneApply<- (CisConfig-class), 13
- geneApply<- ,CisConfig, function-method  
(CisConfig-class), 13
- geneIndcol (transManager-class), 31
- geneNames (transManager-class), 31
- genesym, 24
- GenomicRanges, 15
- GenomicRangesORGRangesList, 15
- GenomicRangesORmissing, 15
- getAll (best.cis.eQTLs), 6
- getBest (best.cis.eQTLs), 6
- getCall (best.cis.eQTLs), 6
- getCisMap, 8, 22
- getSS, 3, 7, 9, 10, 12, 13, 26, 33
- GGtools (GGtools-package), 2
- GGtools-package, 2
- GRanges, 15
- gwSnpScreenResult-class (gwSnpTests), 24
- gwSnpTests, 24
- gwSnpTests, formula, smlSet, missing-method  
(gwSnpTests), 24
- gwSnpTests, formula, smlSet-method  
(gwSnpTests), 24
  
- initialize (CisConfig-class), 13
- initialize, CisConfig-method  
(CisConfig-class), 13
  
- keepMapCache (CisConfig-class), 13
- keepMapCache, CisConfig-method  
(CisConfig-class), 13
- keepMapCache<- (CisConfig-class), 13
- keepMapCache<- ,CisConfig, logical-method  
(CisConfig-class), 13
  
- locusNames (transManager-class), 31
  
- mcwAllCis-class (All.cis), 3
- mcwBestCis, 8, 9
- mcwBestCis-class (best.cis.eQTLs), 6
- meqtlTests (eqtlTests), 18
- meta.All.cis.eQTLs (best.cis.eQTLs), 6
- meta.best.cis.eQTLs (best.cis.eQTLs), 6
- meta.bindmaf (bindmaf), 12
- meta.richNull (richNull), 26
- meta.transScores (transScores), 32
- mtransScores (transScores), 32
  
- nperm (CisConfig-class), 13
- nperm, CisConfig-method  
(CisConfig-class), 13
- nperm<- (CisConfig-class), 13
- nperm<- ,CisConfig, integer-method  
(CisConfig-class), 13
- nthScores (transManager-class), 31
  
- pifdr, 16, 25
- plot, gwSnpScreenResult, character-method  
(gwSnpTests), 24
- probeId, 24

- probesManaged (eqtlTestsManager-class),  
19
- radius (CisConfig-class), 13
- radius, CisConfig-method  
(CisConfig-class), 13
- radius<- (CisConfig-class), 13
- radius<-, CisConfig, integer-method  
(CisConfig-class), 13
- regressOut, 10
- rhs (CisConfig-class), 13
- rhs, CisConfig-method (CisConfig-class),  
13
- rhs<- (CisConfig-class), 13
- rhs<-, CisConfig, formula-method  
(CisConfig-class), 13
- rhs<-, CisConfig, function-method  
(CisConfig-class), 13
- richNull, 26
- schrpref (CisConfig-class), 13
- schrpref, CisConfig-method  
(CisConfig-class), 13
- schrpref<- (CisConfig-class), 13
- schrpref<-, CisConfig, character-method  
(CisConfig-class), 13
- sensanal, 27
- sensanal, sensiCisInput, numeric-method  
(sensiCisInput-class), 28
- sensiCisInput-class, 28
- sensiCisOutput-class, 29
- shortfac (CisConfig-class), 13
- shortfac, CisConfig-method  
(CisConfig-class), 13
- shortfac<- (CisConfig-class), 13
- shortfac<-, CisConfig, integer-method  
(CisConfig-class), 13
- show (CisConfig-class), 13
- show, allCigCis-method (best.cis.eQTLs),  
6
- show, allSigCis-method (best.cis.eQTLs),  
6
- show, CisConfig-method  
(CisConfig-class), 13
- show, cisMap-method (getCisMap), 22
- show, cwBestCis-method (best.cis.eQTLs),  
6
- show, eqtlTestsManager-method  
(eqtlTestsManager-class), 19
- show, gwSnpScreenResult, character-method  
(gwSnpTests), 24
- show, gwSnpScreenResult-method  
(gwSnpTests), 24
- show, mcwAllCis-method (All.cis), 3
- show, mcwBestCis-method  
(best.cis.eQTLs), 6
- show, metaVCF-method (vcf2sm), 35
- show, sensiCisInput-method  
(sensiCisInput-class), 28
- show, sensiCisOutput-method  
(sensiCisOutput-class), 29
- show, transManager-method  
(transManager-class), 31
- smchrpref (CisConfig-class), 13
- smchrpref, CisConfig-method  
(CisConfig-class), 13
- smchrpref<- (CisConfig-class), 13
- smchrpref<-, CisConfig, character-method  
(CisConfig-class), 13
- smFilter (CisConfig-class), 13
- smFilter, CisConfig-method  
(CisConfig-class), 13
- smFilter<- (CisConfig-class), 13
- smFilter<-, CisConfig, function-method  
(CisConfig-class), 13
- smlSet, 18
- smpack (CisConfig-class), 13
- smpack, CisConfig-method  
(CisConfig-class), 13
- smpack<- (CisConfig-class), 13
- smpack<-, CisConfig, character-method  
(CisConfig-class), 13
- snp.rhs.tests, 8, 10, 13, 18
- snpannopk (CisConfig-class), 13
- snpannopk, CisConfig-method  
(CisConfig-class), 13
- snpannopk<- (CisConfig-class), 13
- snpannopk<-, CisConfig, character-method  
(CisConfig-class), 13
- snplocsDefault, 30
- snpsManaged (eqtlTestsManager-class), 19
- SSgen (CisConfig-class), 13
- SSgen, CisConfig-method  
(CisConfig-class), 13
- SSgen<- (CisConfig-class), 13
- SSgen<-, CisConfig, function-method  
(CisConfig-class), 13

strMultPop, [30](#)

topFeats (eqtlTestsManager-class), [19](#)  
topFeats, probeId, eqtlTestsManager-method  
    (eqtlTestsManager-class), [19](#)  
topFeats, rsid, eqtlTestsManager-method  
    (eqtlTestsManager-class), [19](#)  
topGenes (transManager-class), [31](#)  
topScores (transManager-class), [31](#)  
topSnps (gwSnpTests), [24](#)  
topSnps, gwSnpScreenResult-method  
    (gwSnpTests), [24](#)  
tr1\_obs (transScores), [32](#)  
tr1\_perm (transScores), [32](#)  
transManager-class, [31](#)  
transScores, [32](#)  
transTab, [31](#), [34](#)  
transTab, transManager, character-method  
    (transTab), [34](#)  
transTab, transManager, missing-method  
    (transTab), [34](#)

vcf2sm, [35](#)  
vcf2sm, TabixFile, GRanges, integer-method  
    (vcf2sm), [35](#)

Vector, [15](#)