

dsQTL: exploring DNA-variants associated with DNaseI hypersensitivity

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1 Introduction

Degner et al. (2012) publish information on associations between DNA variants (SNP, SNV, and indels) and DNaseI hypersensitivity measures acquired via DNase-Seq.

This package includes information from the Chicago group on normalized DNase-seq data for chromosomes 2 and 17, and genotype data from chromosome 2 only.

2 The basic data structure

```
> library(dsQTL)
> data(DSQ_17)
> exptData(DSQ_17)
```

```
SimpleList of length 2
names(2): MIAME annotation
```

```
> exptData(DSQ_17)[[1]]
```

Experiment data

Experimenter name: Degner JF

Laboratory: Department of Human Genetics, University of Chicago, Chicago, Illinois 60

Contact information:

Title: DNaseI sensitivity QTLs are a major determinant of human expression variation.

URL:

PMIDs: 22307276

Abstract: A 252 word abstract is available. Use 'abstract' method.

We use summarized experiment structure for the assay data, but the imputed genotype data are kept separate, in the package, in the inst/parts folder.

The data structure on chr2, which will be used to reproduce some findings, is more mature

```
> data(DSQ_2)
> names(assays(DSQ_2))
```

```
[1] "normDHS"
```

```
> assays(DSQ_2)[[1]][1:5,1:5]
```

	NA18486	NA18498	NA18499	NA18501	NA18502
dhs_2_1202	-0.2684343	-0.78076674	-0.4840237	2.3894003	-1.0813642
dhs_2_1602	-1.4445813	0.92170439	0.5812017	0.8627376	0.5186581
dhs_2_2002	0.7624075	-0.12340745	-1.1821308	1.4253179	0.3125592
dhs_2_7502	0.1242963	0.60788505	0.6754706	-0.0452303	0.4876332
dhs_2_8802	-0.9554503	-0.06016578	-0.1990696	1.9383937	-1.3758668

```
> rowData(DSQ_2)
```

GRanges with 96024 ranges and 0 elementMetadata cols:

	seqnames	ranges	strand
	<Rle>	<IRanges>	<Rle>
dhs_2_1202	chr2	[1202, 1301]	*
dhs_2_1602	chr2	[1602, 1701]	*
dhs_2_2002	chr2	[2002, 2101]	*
dhs_2_7502	chr2	[7502, 7601]	*
dhs_2_8802	chr2	[8802, 8901]	*
dhs_2_14202	chr2	[14202, 14301]	*
dhs_2_14302	chr2	[14302, 14401]	*
dhs_2_34902	chr2	[34902, 35001]	*
dhs_2_35102	chr2	[35102, 35201]	*
...
dhs_2_242689402	chr2	[242689402, 242689501]	*
dhs_2_242689502	chr2	[242689502, 242689601]	*
dhs_2_242696902	chr2	[242696902, 242697001]	*
dhs_2_242697402	chr2	[242697402, 242697501]	*
dhs_2_242698102	chr2	[242698102, 242698201]	*
dhs_2_242711702	chr2	[242711702, 242711801]	*
dhs_2_242737502	chr2	[242737502, 242737601]	*
dhs_2_242737902	chr2	[242737902, 242738001]	*
dhs_2_242739902	chr2	[242739902, 242740001]	*

```
---
seqlengths:
  chr2
  NA
```

To implement the GGBase protocol for on-the-fly generation of `smlSet` instances from `getSS` queries, we have an `ExpressionSet` instance with specific names.

```
> data(eset, package="dsQTL")
> ex

ExpressionSet (storageMode: lockedEnvironment)
assayData: 96024 features, 70 samples
  element names: exprs
protocolData: none
phenoData
  sampleNames: NA18486 NA18498 ... NA19257 (70 total)
  varLabels: naid one ... isFounder (9 total)
  varMetadata: labelDescription
featureData: none
experimentData: use 'experimentData(object)'
Annotation:
```

The genotype data supplied by Degner et al are imputed to 1000 genomes haplotypes, and are reals in $[0,2]$. For simplicity the current image of the data uses the rounding of the fractional genotypes x with `round(x,0)`.

The feature data refer to the retained 100bp segments that were summarized for DNaseI hypersensitivity and found to lie in the uppermost 5% of the distribution.

```
> library(Biobase)
> fData(ex)[1:5, , drop=FALSE]

data frame with 0 columns and 5 rows
```

We can get the integrated container as

```
> library(GGBase)
> ds2 = getSS("dsQTL", "roundGT_2")
```

the name indicates that we simply rounded the imputed fractional genotypes to nearest integer.

A very restricted search is:

```

> # need to get rid of SNPlocs package getSNPlocs
> getSNPlocs = dsQTL::getSNPlocs # force
> library(GGtools)
> #library(parallel)
> #options(mc.cores=12)
> n1 = best.cis.eQTLs(smpack="dsQTL", radius=2000, geneannopk="dsQTL",
+   snpannopk="dsQTL", chrnames="2", smchrpref="roundGT_",
+   smFilter = function(x) GTFfilter(x, lower=0.05)[23810:23830,],
+   # geneApply=mclapply)
+   geneApply=lapply)

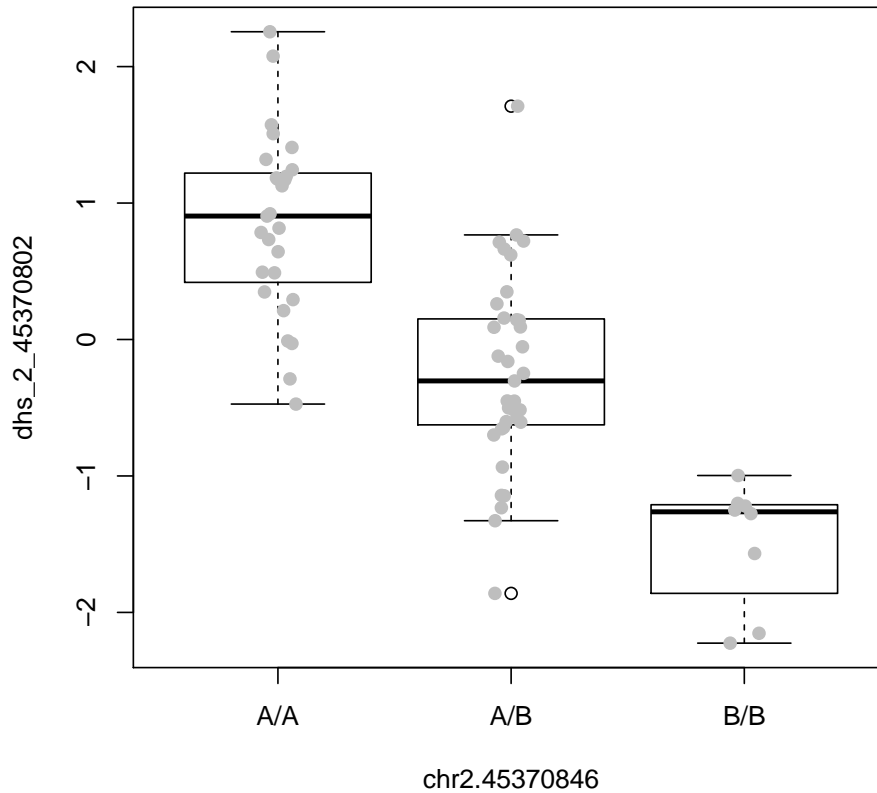
get data...build map...run smFilter...filter probes in map...tests...filter...done.
get data...build map...run smFilter...filter probes in map...tests...filter...done.
get data...build map...run smFilter...filter probes in map...tests...filter...done.

> n1

GGtools mcwBestCis instance. The call was:
best.cis.eQTLs(smpack = "dsQTL", radius = 2000, chrnames = "2",
  smchrpref = "roundGT_", geneApply = lapply, geneannopk = "dsQTL",
  snpannopk = "dsQTL", smFilter = function(x) GTFfilter(x,
    lower = 0.05)[23810:23830, ])
Best loci for 21 are recorded.
Top 4 probe:SNP combinations:
GRanges with 4 ranges and 5 elementMetadata cols:
      seqnames          ranges strand |      score      snpid
      <Rle>            <IRanges> <Rle> | <numeric> <character>
dhs_2_45370802         2 [45368802, 45372901] * |    38.64 chr2.45370846
dhs_2_45370702         2 [45368702, 45372801] * |    29.11 chr2.45370846
dhs_2_45369802         2 [45367802, 45371901] * |    19.14 chr2.45370846
dhs_2_45305002         2 [45303002, 45307101] * |     6.43 chr2.45307016
      snploc radiusUsed      fdr
      <integer> <numeric> <numeric>
dhs_2_45370802 45370846      2000 0.0000000
dhs_2_45370702 45370846      2000 0.0000000
dhs_2_45369802 45370846      2000 0.0000000
dhs_2_45305002 45307016      2000 0.1666667
---
seqlengths:
  2
NA
====
use chromsUsed(), fullreport(), etc. for additional information.

```

```
> plot_EvG(probeId("dhs_2_45370802"), rsid("chr2.45370846"), getSS("dsQTL", "roundGT_
```



3 Session information

```
> sessionInfo()
```

R version 2.15.1 (2012-06-22)

Platform: x86_64-unknown-linux-gnu (64-bit)

locale:

[1] LC_CTYPE=en_US.UTF-8	LC_NUMERIC=C
[3] LC_TIME=en_US.UTF-8	LC_COLLATE=C
[5] LC_MONETARY=en_US.UTF-8	LC_MESSAGES=en_US.UTF-8
[7] LC_PAPER=C	LC_NAME=C
[9] LC_ADDRESS=C	LC_TELEPHONE=C
[11] LC_MEASUREMENT=en_US.UTF-8	LC_IDENTIFICATION=C

attached base packages:

```
[1] stats4    splines  stats    graphics grDevices utils    datasets
[8] methods  base
```

other attached packages:

```
[1] GGtools_4.4.0      Rsamtools_1.8.5    Biostrings_2.24.1
[4] dsQTL_0.0.18      GGBase_3.18.0      snpStats_1.6.0
[7] Matrix_1.0-6      lattice_0.20-6     survival_2.36-14
[10] Biobase_2.16.0    GenomicRanges_1.8.7 IRanges_1.14.4
[13] BiocGenerics_0.2.0
```

loaded via a namespace (and not attached):

```
[1] AnnotationDbi_1.18.1  BSgenome_1.24.0    DBI_0.2-5
[4] GenomicFeatures_1.8.2 RCurl_1.91-1        RSQLite_0.11.1
[7] VariantAnnotation_1.2.9 XML_3.9-4           annotate_1.34.1
[10] biomaRt_2.12.0       bit_1.1-8           bitops_1.0-4.1
[13] ff_2.2-7             genefilter_1.38.0  grid_2.15.1
[16] rtracklayer_1.16.2   tools_2.15.1       xtable_1.7-0
[19] zlibbioc_1.2.0
```